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1.0 Introduction

Natural products containing conjugated alkenyl units represent a large and structurally diverse group of compounds. Particularly significant are those possessing biological activity, including the eicosanoids (the arachidonic acid derivatives such as the leukotrienes and lipoxins),¹ the retinoids,² and the polyene macrolides,³ a large group comprising over 200 members. There are also a considerable number of relatively unexplored polyenes derived from a variety of sources, including marine organisms, fungi, slime-moulds and plants; these polyenes are often produced as a means of chemical defense, and thus also possess potentially useful biological activity.

This review covers the literature from 1971 to the present date. It begins with a brief survey of the major sub-categories of polyene natural products, before focussing on methods used to bring about their synthesis. Heavier emphasis will be given to the synthesis of the polyenes themselves and owing to the vast number of natural products containing multiple double bonds, this review will concentrate only on natural products that contain at least a) three all-*trans* conjugated double bonds, or b) *cis,cis* or *cis,trans* dienyl units.

1.1 Polyenes derived from bacteria

Typically isolated from the *Streptomyces* genera of actinomycetes soil bacteria, the polyene and oxopolyene macrolides⁴ are relations of the more familiar macrolide antibiotics (for example, the erythromycins) and represent the most important sub-category of polyene natural products. Structurally, they are very large macrocyclic lactones with 22 to 44 membered rings commonplace. The polyenic section usually incorporates between 4 and 8 conjugated double bonds. Oxopolyene macrolides such as roxaticin⁵ **1** are characterized by having the polyene in conjugation with the lactone linkage (Fig. 1). Several members of the polyene macrolide family find service as clinically important anti-fungal drugs, notably pimarinic acid⁶, nystatin A₁^{3,7} and amphotericin B **4**,⁸ dubbed ‘amphoterrible’ due to its high nephrotoxicity.

The ansamycin⁹ antibiotics are a growing family of *Streptomyces* metabolites, and many of them possess antibacterial, antifungal or antitumour activity. Ansatrienine A (mycotriene)

5¹⁰ is one of the first isolated examples, and may be taken as a representative member. Also isolated from *Streptomyces* sp., the manumycins^{11,12} (for example, **6** and **7**) are a fairly large and well-studied group of polyenes. This family of antibiotics has attracted considerable attention due to their promising biological activity; manumycin A **6**, the first to be isolated, shows activity towards fungi and L1210 leukemia stem cells in addition to its antibacterial properties.

Isolated in 1994 from the fermentation broth of the gliding bacteria *Sorangium cellulosum*, the disorazoles,¹³ of which disorazole F₂ **8** is typical, are a group of 29 anti-fungal and cytotoxic macrocyclic dilactones. They are all dimer-like, and are differentiated by the positions and stereochemistries of the double bonds, and in the size of, and nature of the substituents on the macrocyclic core.

Aside from these major classes, there are a host of other bacterially derived polyenes (Fig 2). Xanthomonadin I **9** responsible for the vivid yellow colours of *Xanthomonas* colonies, is a rare example of a compound synthesized by a terrestrial organism that contains bromine.¹⁴ Linearmycin A₁ **10**¹⁵ is an unusually long polyene containing thirteen double bonds, whilst the structurally complex polyenes viridenomycin **11**¹⁶ (an antifungal, antibacterial and anti-tumour macrolide) and rapamycin **12**¹⁷ (an antibacterial and immunosuppressive agent) are further examples of *Streptomyces* metabolites.

1.2 Polyenes produced by fungi

Pigments derived from fungi tend to be of isoprenoidal origin, not arising from the polyketide metabolic pathways that usually give rise to polyenic compounds. There are a few examples however, and a prominent one is that of fumigillin **13**^{18,19} (Fig. 3), a metabolite of *Aspergillus fumigatus* that possesses a wealth of biological activities including amebicidal, anticancer, antiparasitic and antibacterial properties. Also important are the rhizoxins,²⁰ a family of 16-membered macrolactones isolated from the plant-pathogenic fungus *Rhizopus chinensis*. Rhizoxin **14** is a tubulin binding anti-mitotic agent possessing antifungal and antimicrobial activity in addition to potent *in vitro* cytotoxicity and *in vivo* antitumour activity. It is more potent, yet less toxic than vincristine and has undergone extensive clinical trials. The dideseoxy analogue rhizoxin D²¹ has equal potency, but is isolated as a minor component.

1.3 Polyenes derived from slime-moulds and plant sources

Slime moulds, often found amongst moist decaying wood and litter in forests, produce yellow cytotoxic substances such as fuligrobin A **15**²² and physarochrome A **16**²³ (Fig. 4), either as a means of chemical defense, or to function as photoreceptors. Plants synthesize a vast array of polyenic compounds, most familiar of which are the carotenoids²⁴ (e.g. γ -carotene **17**), the most prevalent of all the naturally occurring pigments (Fig 5). The deoxyphorbol ester derivatives **18** and **19** isolated from the caustic sap of the pencil tree, are purported anti-tumour agents.²⁵ Also shown are several unusual polyenes containing mixed alkene geometries **20–22**.²⁶

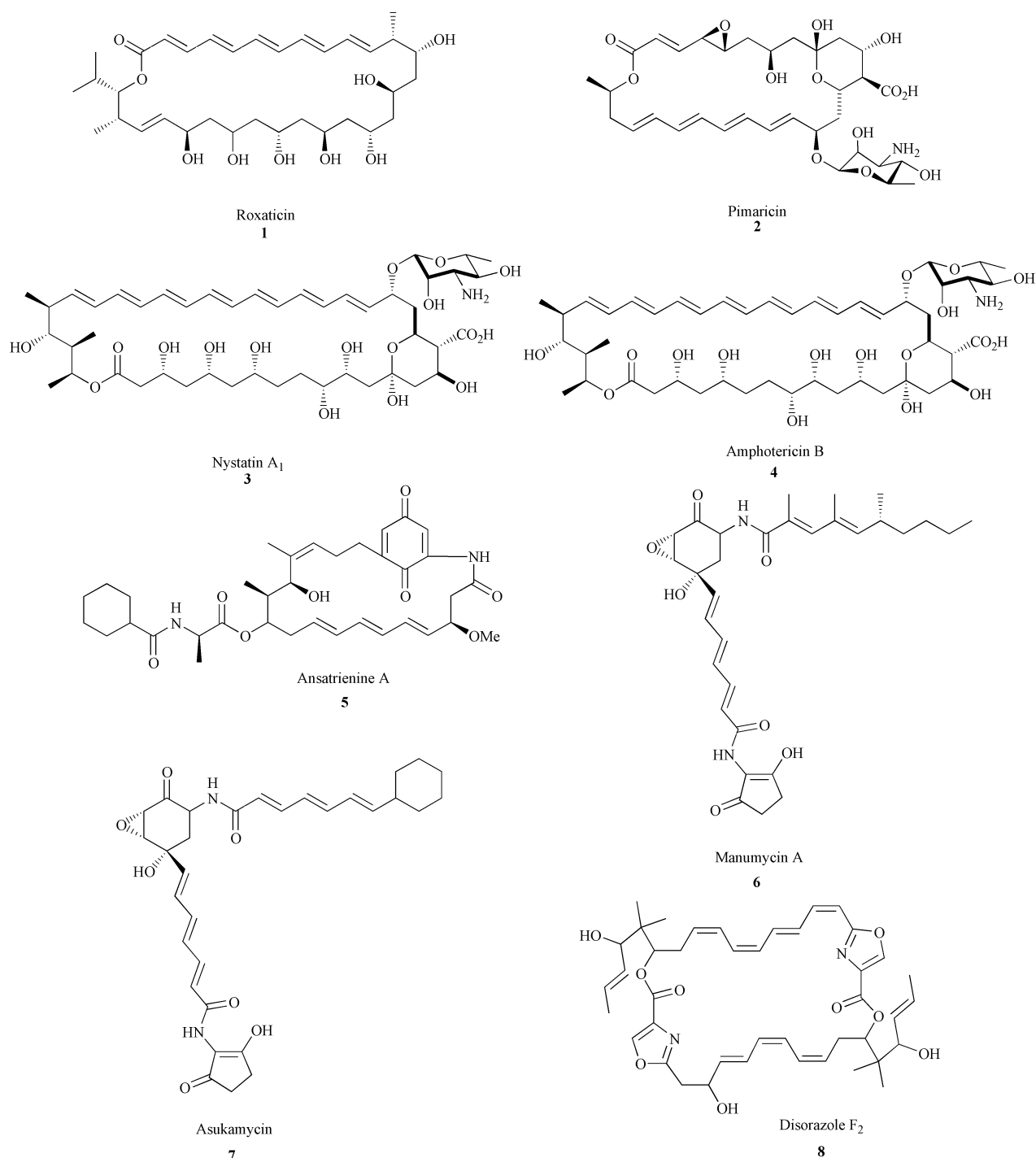


Fig. 1

1.4 Polyenes derived from marine organisms

Naturally derived polyenes are typically toxic, and often vividly coloured. These two facets are exploited well by many marine organisms which are without the defensive capabilities of many of their larger aquatic cousins. Opisthobranchia for example, a sub-class comprising the sea-slugs and their allies, lack the protection provided by the hard shell typical of most members of the gastropod class, and secrete brightly coloured polyenic ketones, such as the yellow navanones A–C **23–25** (named after *Navanax inermis*, an ornately coloured sea-slug found off the Californian coast) which act as alarm pheromones.²⁷ The cytotoxic aurantosides A and B (**26** and **27**, Fig. 5), obtained from sponges and isolable as orange amorphous powders, are *N*-trisaccharide tetramic acid derivatives possessing activity against P388 and L1210 leukemia cells.^{28a} Aurantosides C^{28b}

and D–F^{28c} have also been recently isolated and found to possess biological activity. The antibiotic keronopsins **28–31** are produced by *Pseudokeronopsis rubra*, a marine ciliate known for its deep red colour, and are used as chemical weapons against other ciliates and flagellates.²⁹ Marine dinoflagellates, a type of unicellular aquatic organism possessing both animal and plant characteristics, provide a rich source of structurally diverse natural products with highly specific bio-activity. Certain species produce powerful nerve toxins, and it is these toxins that give rise to the so-called red-tides, formed when the creatures reproduce *en mass* or bloom in shallow waters. The genus *Amphidinium* has been exploited as a source of novel secondary metabolites with unique chemical structures,³⁰ and amphidinol 3 **32** (Fig 6), an antifungal polyene whose structure was recently elucidated,³¹ is a prominent example.

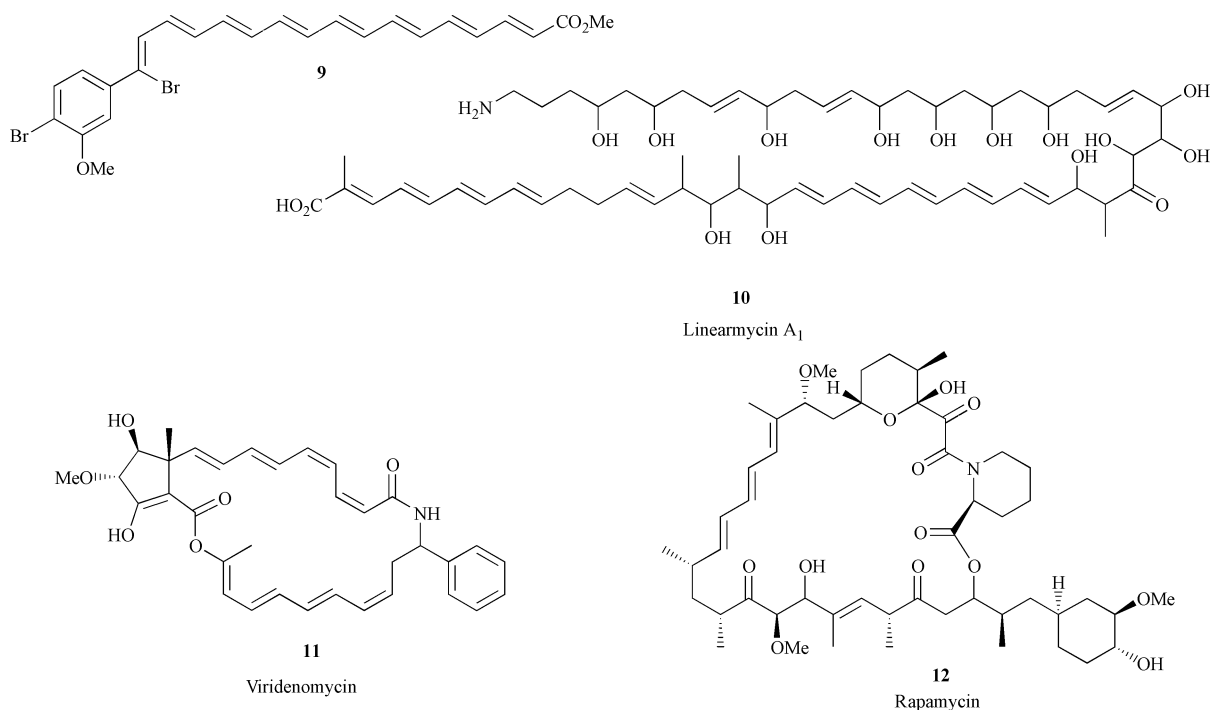
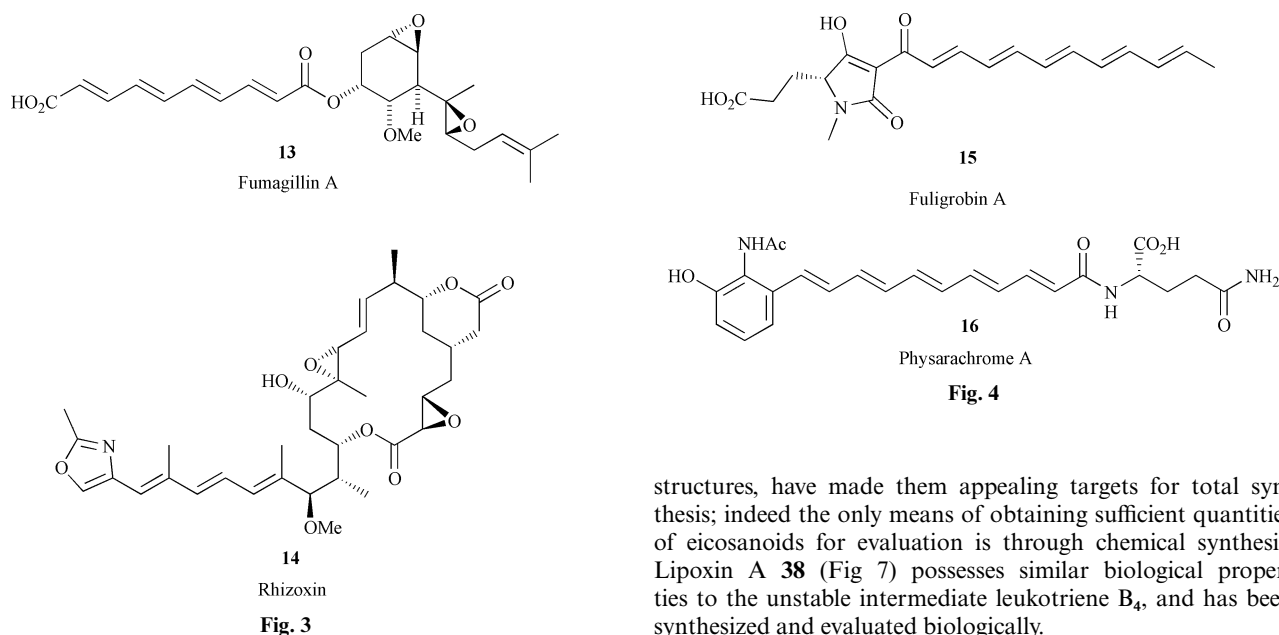


Fig. 2



structures, have made them appealing targets for total synthesis; indeed the only means of obtaining sufficient quantities of eicosanoids for evaluation is through chemical synthesis. Lipoxin A **38** (Fig 7) possesses similar biological properties to the unstable intermediate leukotriene B₄, and has been synthesized and evaluated biologically.

1.5 Polyenes produced by animals

The retinoids² **33–37** (Fig. 7), a collective term for the various synthetic and natural analogs of retinol (vitamin A) **33**, are vitally important during embryo development and throughout post-natal life in humans. The biological roles they play are dependent both on the nature of the R group and on the geometry of the polyene chain. These factors are the reason that they have been the focus of much synthetic attention, and are often used as the first test of a new polyene-assembling methodology.

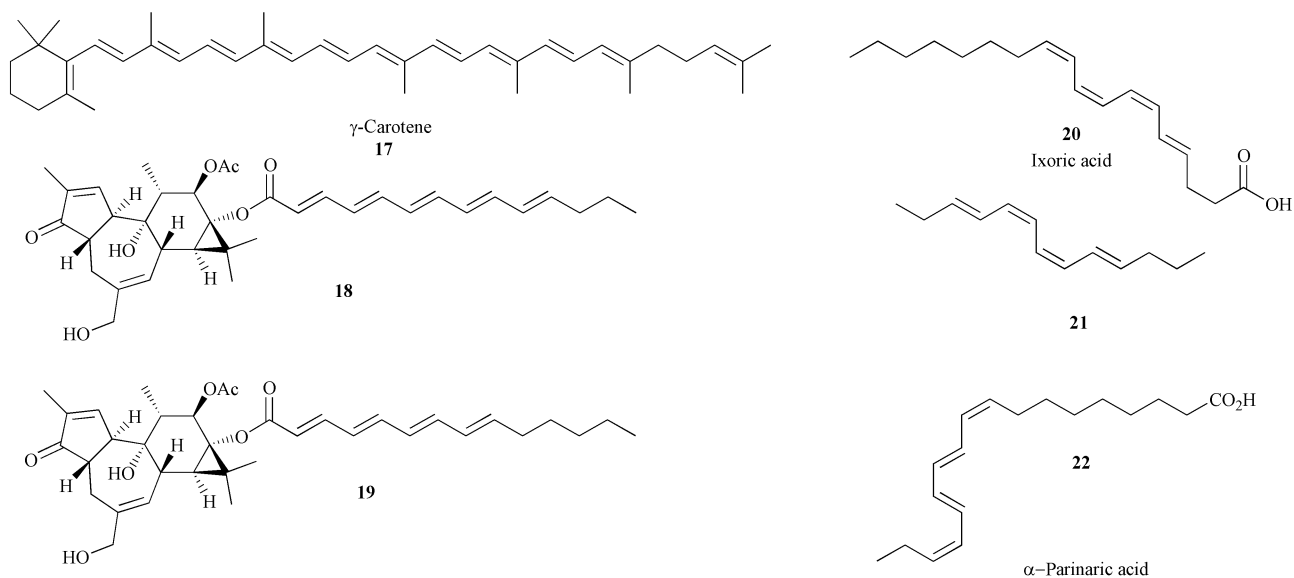
Another important group of polyenic biomolecules are the eicosanoids,¹ comprising the structurally related prostaglandins, prostacyclins, thromboxanes and leukotrienes, which are derived from polyunsaturated fatty acids such as arachidonic acid. These compounds are found in minute quantities in most animal cells, and elicit diverse biological responses in most animals. These effects, together with their small, but complex

2.0 Synthesis of polyenes—general strategies

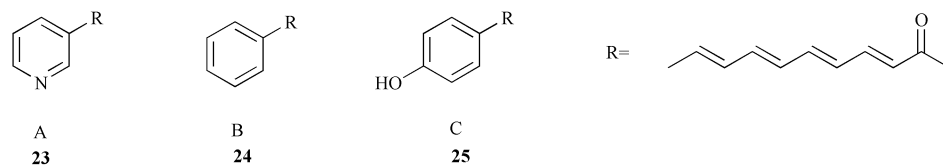
Fig. 8 outlines the most common general strategies taken to date in order to access polyenes.

A few fundamental issues need to be considered when synthesizing polyunsaturated compounds. As a general rule, polyenes of the all-*trans* nature are more stable than those containing *cis*-alkenyl units, hence the latter tend to be more difficult to construct, and especially prone to isomerization. Additionally, when a specific isomer is required, as is usually the case, a method that leads to mixtures of *cis* and *trans* isomers will invariably require a tedious and yield-reducing separation, if separation is possible at all.

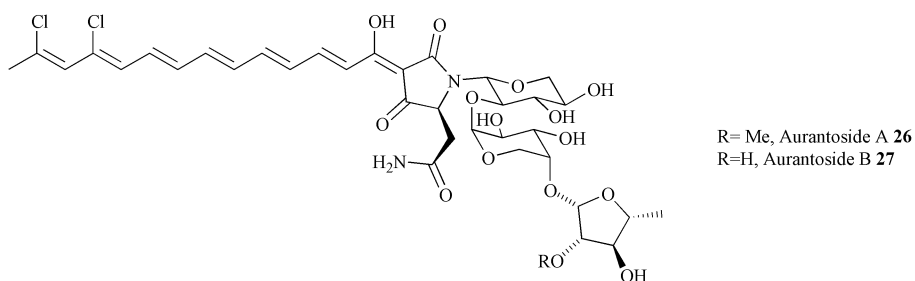
The principal requirements for polyene synthesis are thus 1) a reliable olefination procedure that produces alkenes in high geometric purity, 2) a procedure that allows ready access to either isomer, whilst at the same time being mild and functional group tolerant.



Navanones



Aurantiosides



Keronopsins

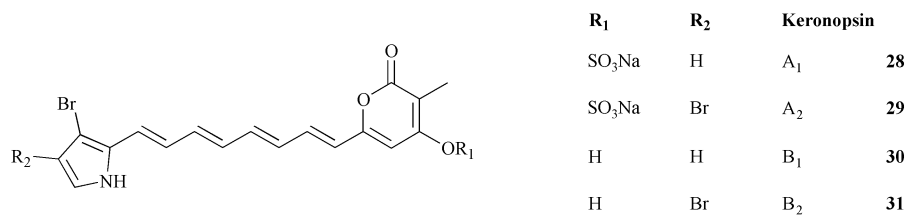


Fig. 5

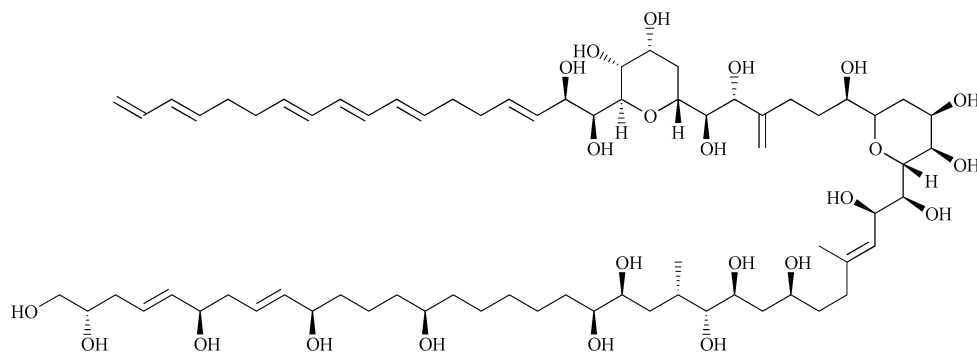
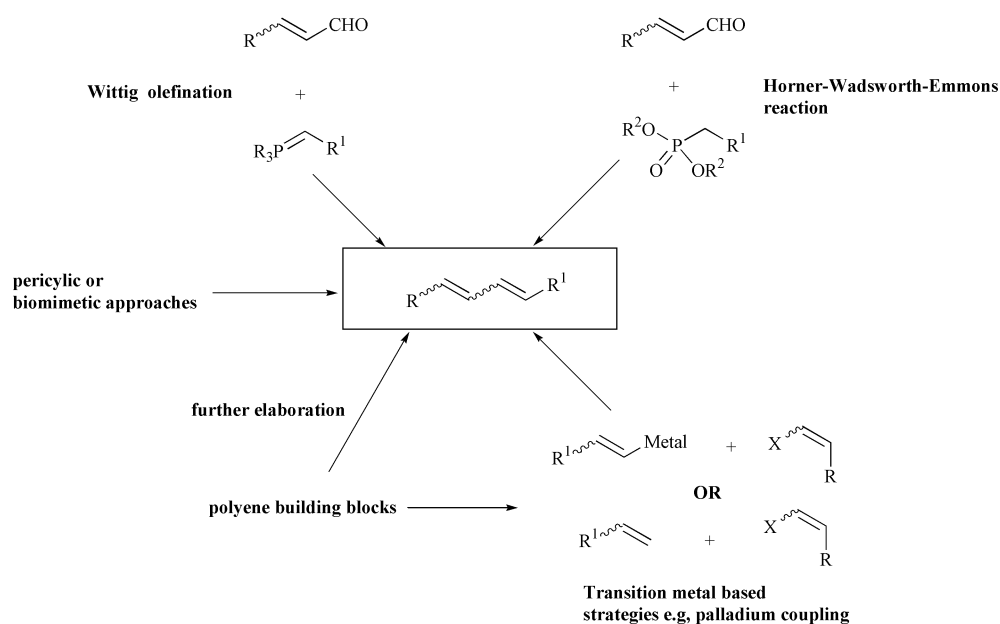
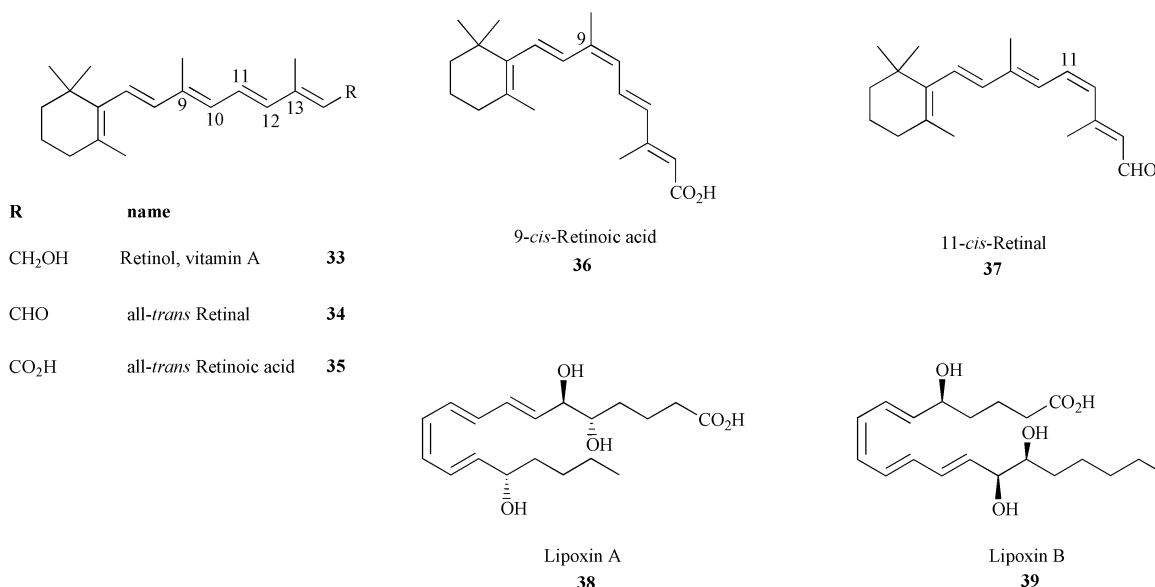


Fig. 6



2.1 Wittig olefination and Horner–Wadsworth–Emmons procedure

The Wittig reaction³² has proved historically to be the reaction of choice for the formation of alkenes. It suffers however from a number of disadvantages, the principal one being that it invariably leads to a mixture of isomers, necessitating separation, or isomerization of the unwanted isomer. It also requires preparation of an appropriate ylide, typically requiring a strong base to generate the carbanion, and usually a laborious purification procedure necessary to remove the stoichiometric phosphine oxide by-product. The closely related Horner–Wadsworth–Emmons³³ (HWE) procedure circumvents a number of these problems, and is now increasingly used in all areas of natural product synthesis.

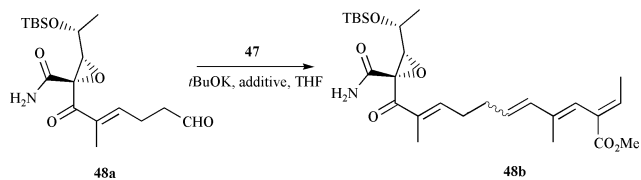
The BASF synthesis of vitamin A, one of the leading methods of production, utilizes Wittig chemistry on a huge scale to produce the precursor retinyl acetate **40** (Scheme 1). A double Wittig reaction was recently employed by Tode's group in the first synthesis of the marine carotenoid crassostreaxanthin **B 41** (Scheme 1).³⁴ Condensation of aldehyde **42** with Wittig salts **43** and **44** led to **41** after deprotection. Wittig chemistry was also employed in the synthesis of the side chain of the

novel neuritogenic agent epolactaene **45** (Scheme 2),³⁵ with the requisite ylide **47** being prepared in six steps starting from known iodoalcohol **46**. Various conditions were examined in order to give the highest *E* : *Z* ratio for the Wittig coupling of **47** and **48** (Table 1); use of the tributylphosphonium salt **47b** gave the highest proportion of the *E* isomer which was separated from the *Z* isomer by flash chromatography.

As mentioned previously, arachidonic acid metabolites are important due to their intriguing biological properties, and Yadav and co-workers have developed a convergent synthesis of lipoxin A **38** (Scheme 3), with Wittig chemistry being used to install the tetraene portion. Their synthesis involves a specialized ylide **49**, prepared from propargyl alcohol (prop-2-yn-1-ol).³⁶ In 1991 Pattenden and Patel reported the total synthesis of all-*trans* citreomontanin **52**, a hexaene isolated from *Penicillium pedemontanum* and the putative precursor to citreoviridin and citreoviridinol. This was achieved *via* an iterative sequence involving Wittig coupling between aldehyde and triphenylphosphoranylpropanoate **50**, propanoate reduction, and oxidation to regenerate an aldehyde **51** (Scheme 4).³⁷

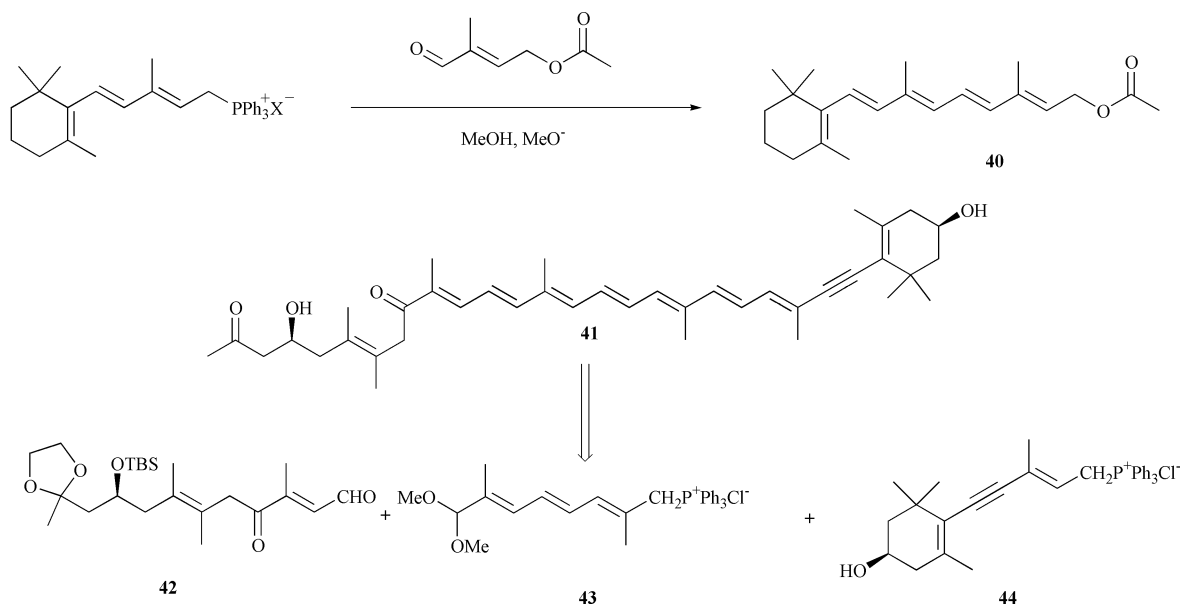
There are numerous applications of the HWE procedure towards polyene synthesis. A prominent example can be seen in Nicolau and co-workers' landmark total synthesis of

Table 1

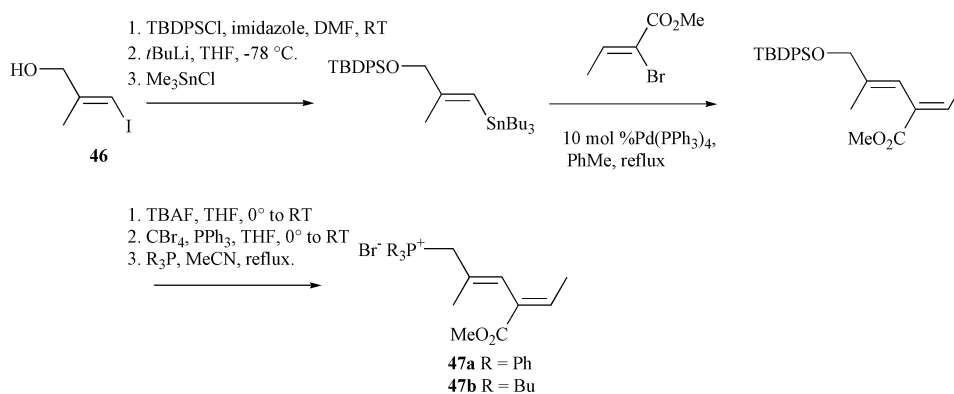
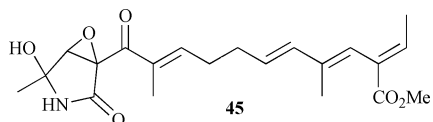


Ylide	Ylide (equivalents)	Additive	<i>T</i> /°C	<i>E</i> : <i>Z</i> Ratio ^a	Yield (%)
47a	2.4	None	-78	—	Trace
47a	2.4	18-Crown-6-MeCN	-46	1 : 1	27
47a	5.0	18-Crown-6-MeCN	-46	1 : 1	68
47b	5.0	18-Crown-6-MeCN	-46	10 : 1	69

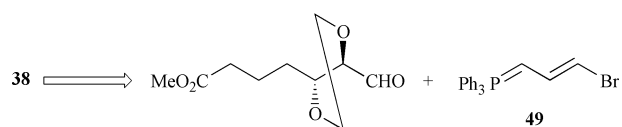
^a Determined by ¹H NMR



Scheme 1

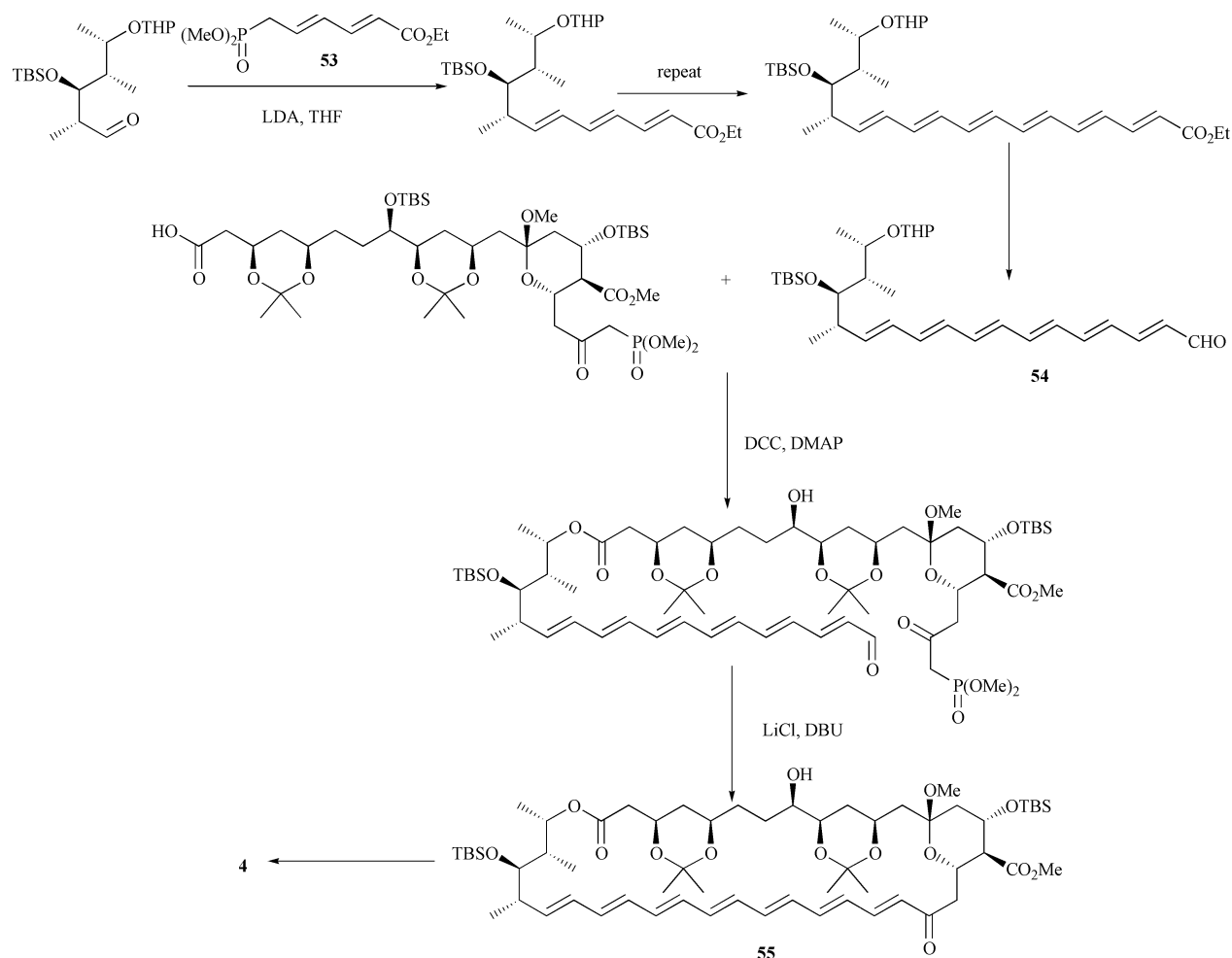
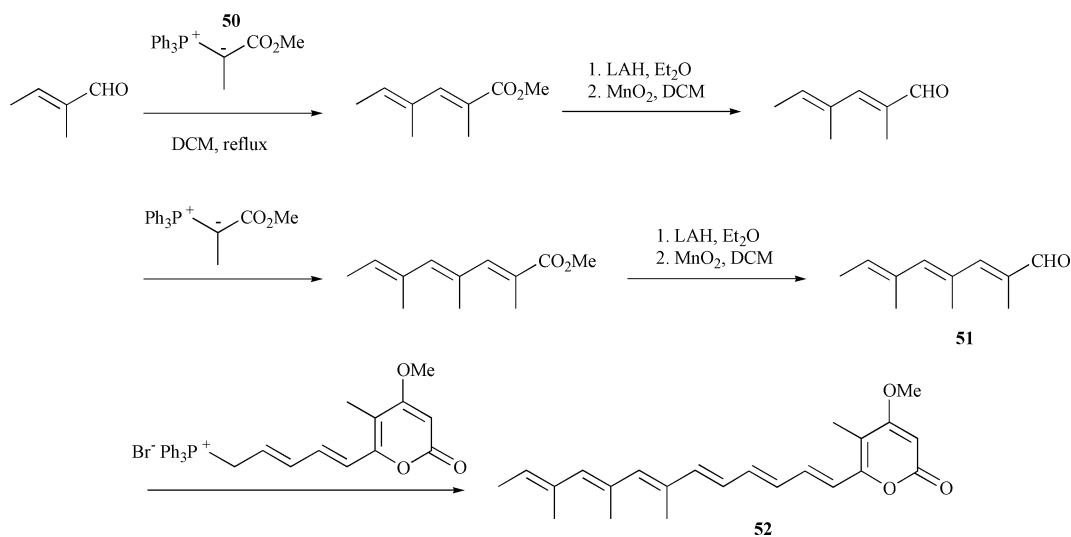


Scheme 2



Scheme 3

amphotericin B **4**, in which three HWE reactions were used to construct the polyene section (Scheme 5).³⁸ The first two employed the functionalized phosphonate **53** leading to the acyclic hexaenal **54**, before the third reaction was used to induce ring closure, giving the cyclic heptaene **55**. A similar approach was taken by Mori's group in their recent synthesis of roxaticin **1**.³⁹

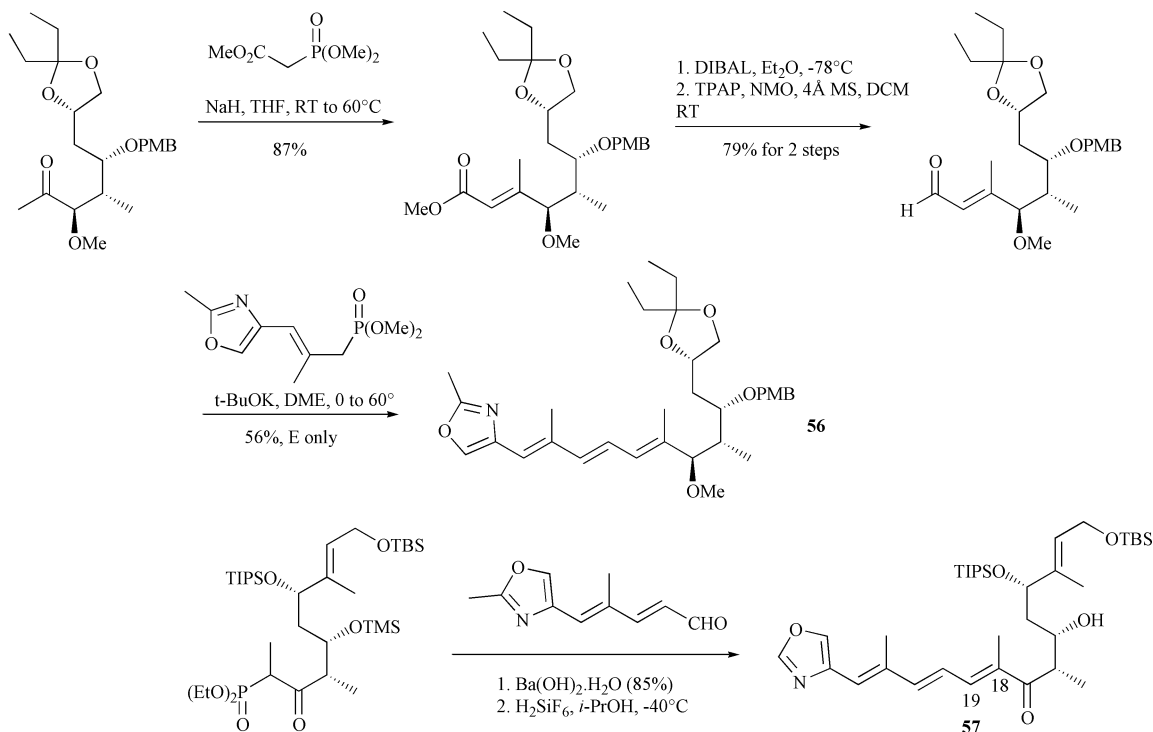


The rhizoxins have attracted considerable attention owing to their remarkable biological activity,²⁰ and many efforts have been directed towards rhizoxin and its congeners.⁴⁰ Burke and co-workers' partial synthesis of rhizoxin^{40e} **14** (Scheme 6) used two HWE reactions to form the trisubstituted *E*-olefin **56**.

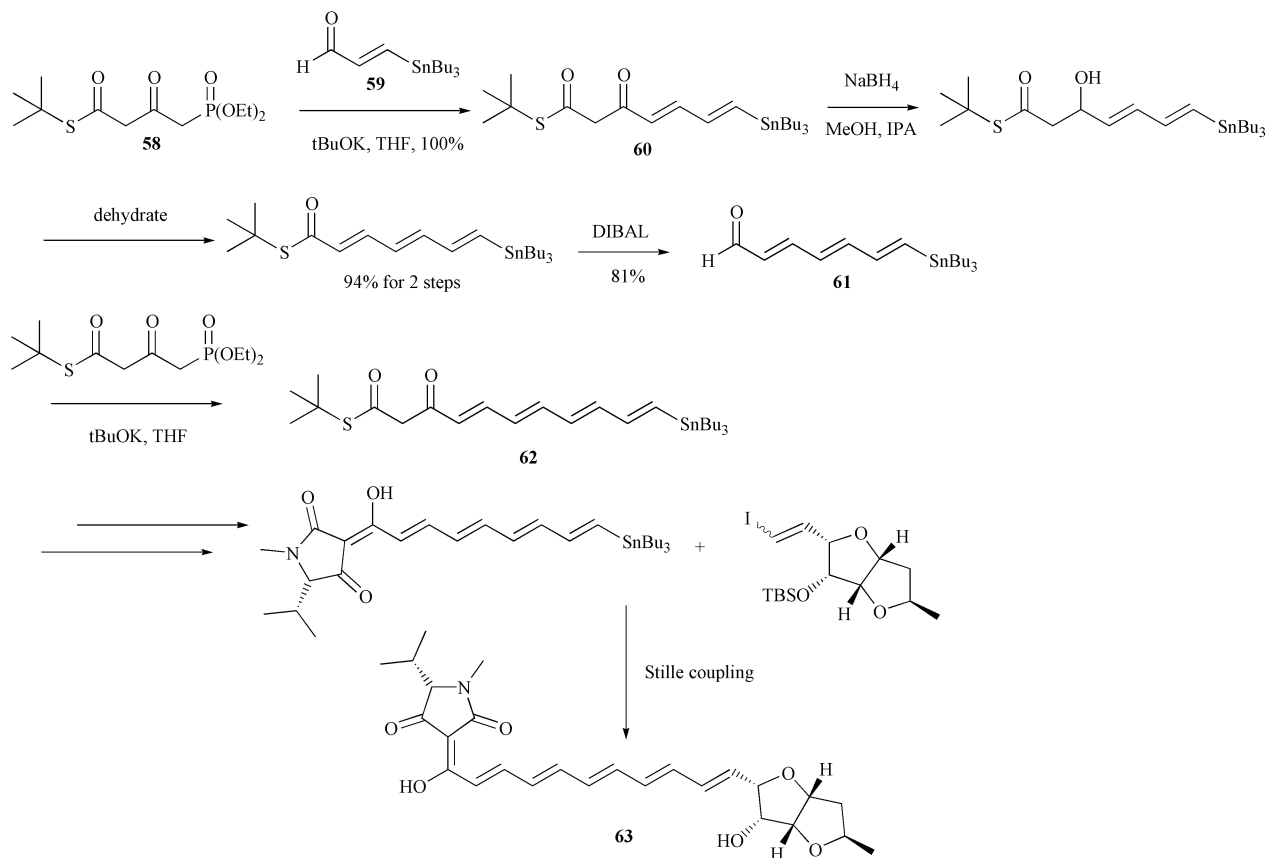
Leahy and co-workers' total synthesis of rhizoxin D utilizes a similar approach.⁴¹ They explored various routes to the tricky triene oxazole fragment **57** before settling on a HWE reaction between C18 and C19. Ley's group used HWE reactions to install the acyltetramic acid portion of the antibiotic erythro-skyrine **63** (Scheme 7).^{42,43} They coupled the phosphonate ester **58** with the β -tributylstannyl aldehyde **59** under optimized

conditions, giving the dienyl stannane **60** quantitatively as a >30 : 1 mixture of *E*-*Z* isomers. Subjecting of trienal **61** and phosphonate **58** to HWE reaction under identical conditions provided the tetraene **62** in high yield and with high selectivity (>30 : 1 *E*-*Z*).

Phosphorous ylide based reactions played a pivotal role in the construction of the macrocyclic lactam cyclamenol A **66** (Scheme 8), recently reported by Nazare and Waldmann.⁴⁴ In devising their synthetic strategy, the authors paid particular attention to the polyene section, which comprised one *Z* and six *E* double bonds. It was feared that the *Z* double bond would isomerize, or the polyene skeleton would react further to give a



Scheme 6

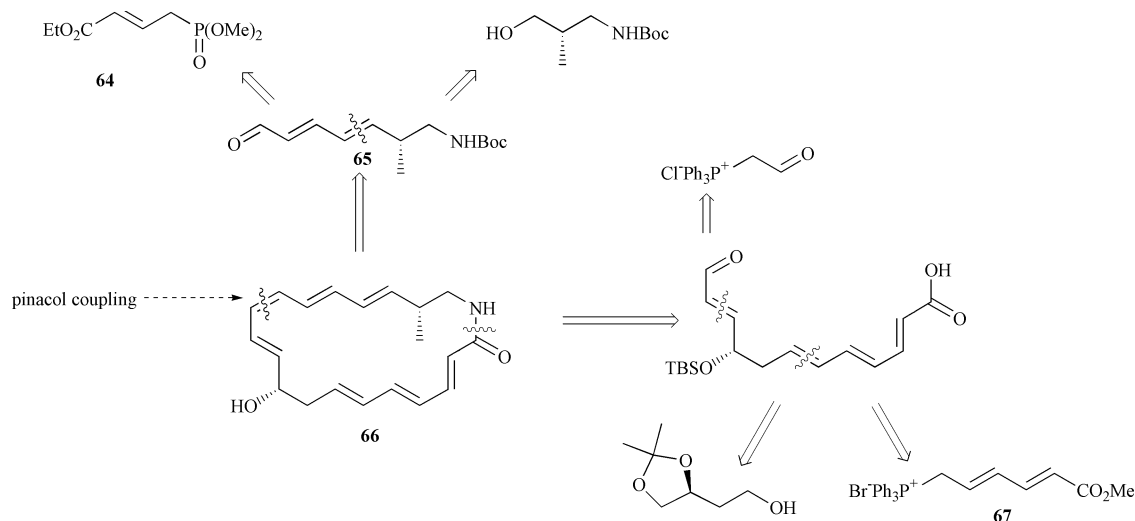


Scheme 7

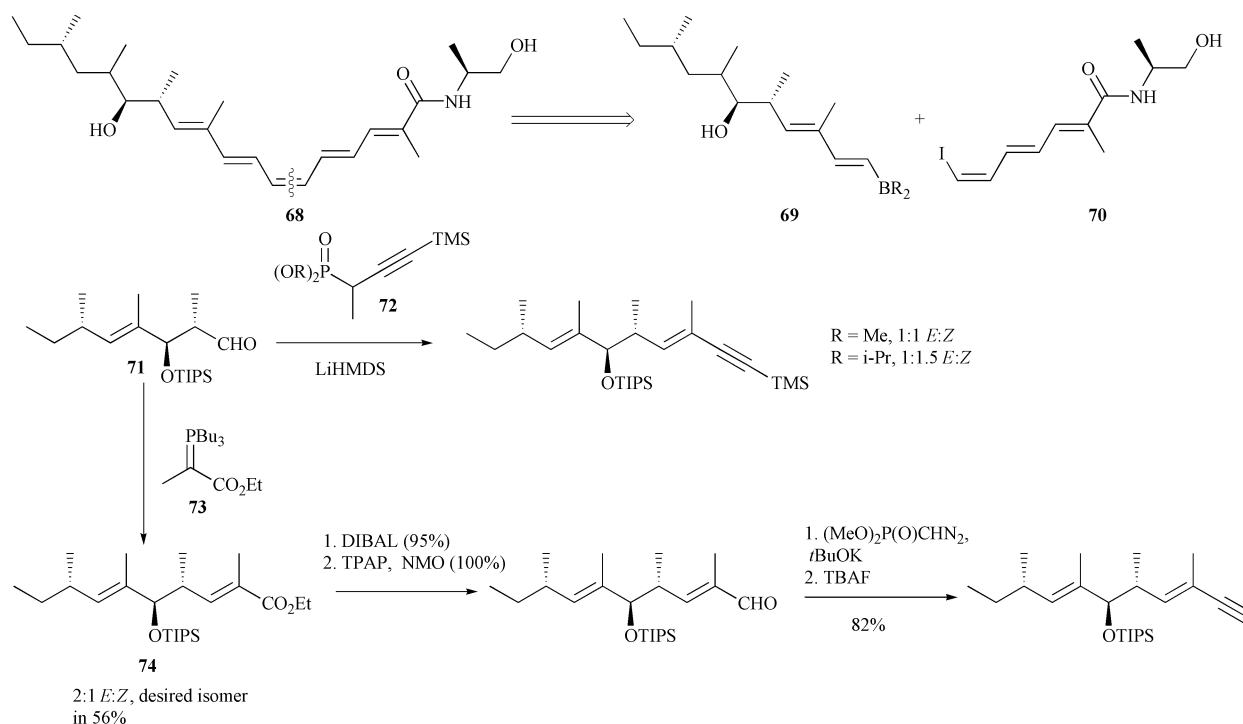
fully conjugated octaene. For this reason, construction of the polyene system was left until late in the synthesis, and the final *Z* double bond was introduced *via* a pinacol coupling, allowing both ends of the acyclic precursor to be tied together through template control. A HWE procedure involving crotonyl phosphonate **64** was used to construct the dienal **65**, whilst a Wittig reaction exploiting the phosphonium salt **67** derived from sorbic acid was used to provide the southern triene.

So far, only examples of the Wittig and HWE reactions lead-

ing to all *E* polyenes have been examined; this is mainly due to the inherent *E*-selectivity of these processes. It is possible however, either by changing the nature of the phosphonium salt/phosphonate or by varying the reaction conditions, to bias the outcome towards either alkene isomer. In the first reported synthesis of myxalamide **A** **68** (Scheme 9), the most abundant of the four myxalamides⁴⁵ isolated from the gliding bacteria *Myxococcus xanthus*, Mapp and Heathcock opted to complete the synthesis *via* a Suzuki coupling (see section 2.2) between



Scheme 8

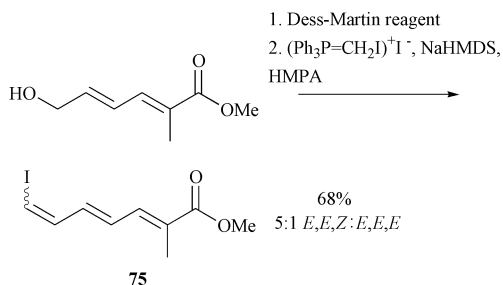


Scheme 9

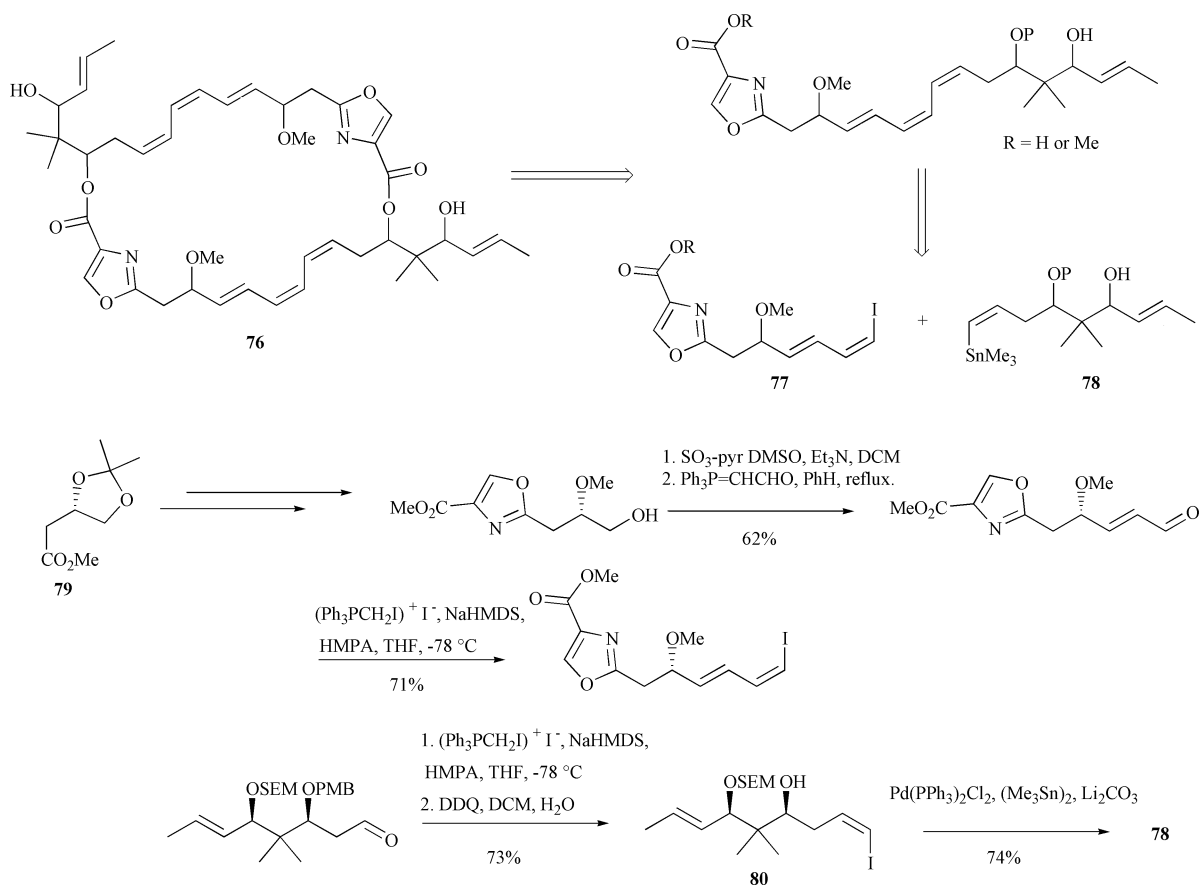
dienylborane **69** and iodotriene **70**.⁴⁶ In order to prepare the enyne precursor that would undergo hydroboration to give **69**, they initially attempted to install the triple bond *via* a HWE reaction between aldehyde **71** and phosphonate **72**. However, *E/Z* selectivities were not good, and use of the bulkier phosphonate gave greater selectivity, but in the wrong direction. Use of the stabilized ylide **73** gave the unsaturated ester **74** in an improved 2 : 1 (*E-Z*) ratio. Synthesis of the triene portion of iodotriene **70** (Scheme 10) required formation of an (*E,E,Z*)-

alkenyl unit **75**, with insertion of the *Z*-alkene being accomplished under kinetic conditions through the Stork–Zhao modification of the Wittig reaction.⁴⁷ Unfortunately, although the ratio of isomers was acceptable, separation by a variety of methods proved impossible, and the authors were forced to use an alternative approach to this fragment.

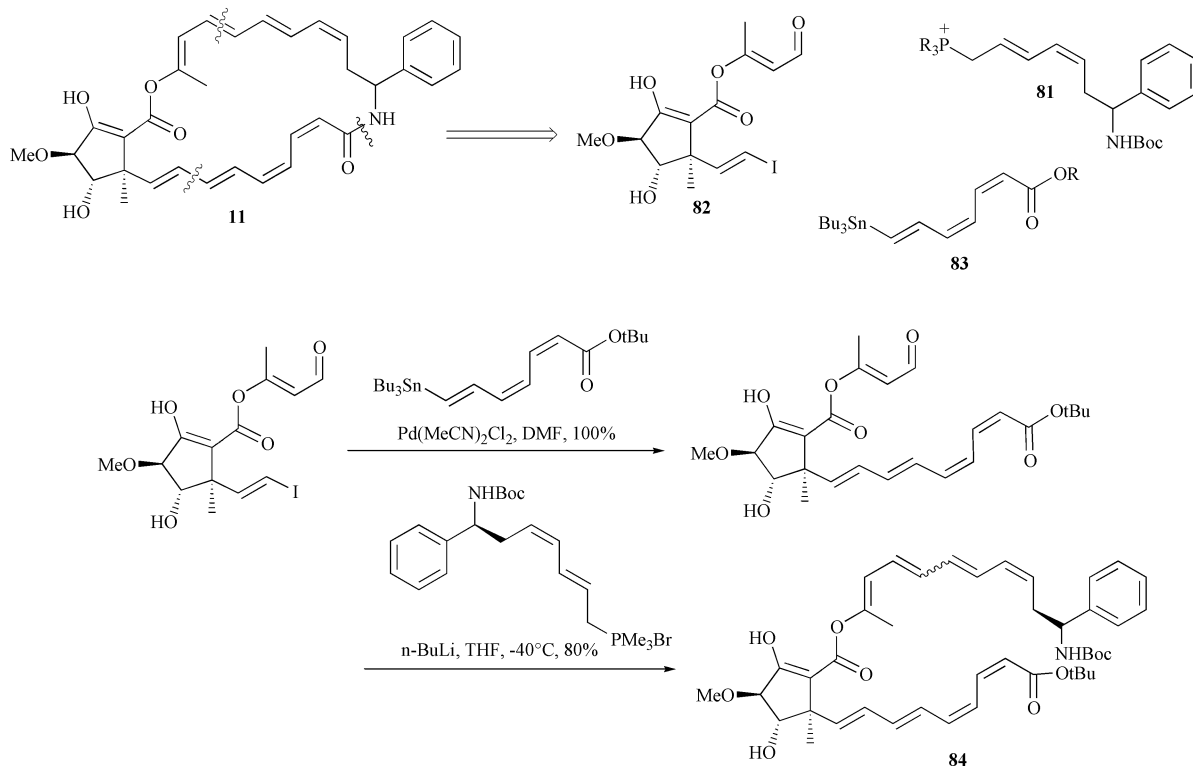
This modification⁴⁷ was also employed in Meyers and co-workers' synthesis of the monomeric unit of diorazole **C**, **76** (Scheme 11).⁴⁸ Here, a palladium coupling reaction was also chosen as the final bond-forming reaction, which retrosynthetically led back to an oxazole containing dienyl iodide **77** and a *cis*-alkenyl stannane **78**. Iodide **77** was assembled from readily available ester **79**, *via* Wittig homologation to insert the *trans* double bond, followed by the Stork–Zhao Wittig modification. This strategy was used again to furnish alkenyl iodide **80**, which gave **78** after the Stille coupling. The same group also recently reported studies towards the total synthesis of viridenomycin⁴⁹ **11** (Scheme 12), a large polyene macrolide containing two mixed geometry tetraenes. In their second generation approach, the authors chose to disconnect **11** into three key fragments, allowing these two potentially problematic units to be synthesized in



Scheme 10



Scheme 11



Scheme 12

isolation. It was envisaged that **81** would be joined to the core cyclopentenol fragment **82** using phosphonate chemistry and **81** was prepared accordingly, with a kinetic Wittig reaction being used to insert the required *Z*-alkene with good selectivity. The original strategy to use a phosphonate to join **81** and **82** failed in studies using model aldehydes (entries 1 and 2, Table 2), as rearrangement to give a ketone was observed. Use of the semi-

stabilized ylide described by Tamura and co-workers⁵⁰ gave poor *E/Z* selectivity (entries 3 and 4). In the end, the authors used trimethylphosphonium salt **81** (R = Me) to prepare **84**, which gave high selectivity towards the desired *E* isomer. A kinetic HWE reaction was employed to synthesize the trienyl stannane **83** (Scheme 13); use of the electrophilic Still–Gennari phosphonate⁵¹ under kinetic conditions that favour rapid

Table 2

Entry	R	PL ₃	Base	Aldehyde ^a	Product ^a	Ratio (E : Z)
1	Me	PO(OEt) ₂	KO <i>t</i> -Bu	85	86	—
2	Me	PO(OEt) ₂	<i>n</i> -BuLi	85	86	—
3	H	PBu ₃	KO <i>t</i> -Bu	87	88	2.6 : 1
4	H	PBu ₃	<i>n</i> -BuLi	87	88	2.9 : 1
5	H	PMe ₃	KO <i>t</i> -Bu	87	88	3.0 : 1
6	H	PMe ₃	<i>n</i> -BuLi	87	88	30 : 1
7	H	PMe ₃	<i>n</i> -BuLi	85	89	10 : 1 (90%)

^a

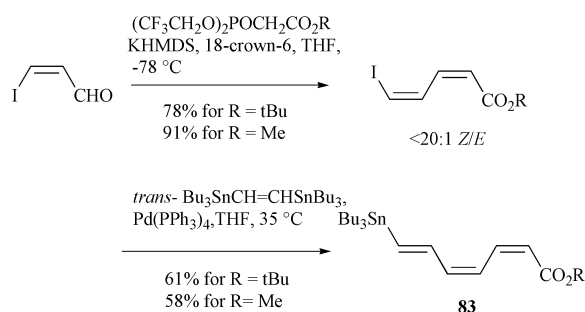
85

86

87

88

89



Scheme 13

elimination leads to *Z*-alkenes in excellent isomeric purity. Unfortunately, removal of both *t*-Bu protecting groups proved impossible, and cyclization of **84** was not realized.

Taylor's group recently disclosed a procedure that enables elaboration of haloenals **90** without the need for their isolation.⁵² This is useful because whilst such compounds are potentially valuable as polyene building blocks, they are also unstable, difficult to isolate, and have a severe irritant effect on mucous membranes, particularly the bromo- and chloro-enals (both isomers of iodopropenal can be prepared readily, but retain lachrymatory properties). Their protocol involves *in situ* manganese dioxide haloenol oxidation, followed by direct Wittig homologation, giving dienyl haloesters **91** that are themselves useful polyene building blocks, given that partial reduction of the ester function would allow the process to be repeated (Scheme 14).

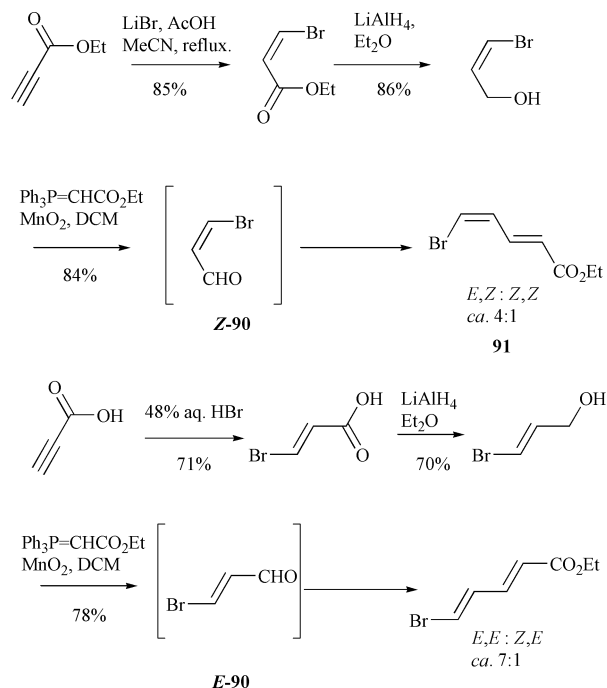
2.2 Transition metal based strategies

Representative of these strategies are the palladium cross-coupling reactions, allowing single bond formation between two sp² centres with excellent stereoselectivity and often under very mild conditions, and without some of the problems encountered with more traditional approaches. The mildness and functional group tolerance of these reactions makes them an ideal way to insert the final 'stitch' in a polyene chain, thus allowing this often troublesome section to be assembled last. The Stille reaction⁵³ is particularly useful, and typically proceeds with retention of configuration of the alkenyl halide, leading to polyenes of exceptionally high geometrical purity. This reaction has been exploited many times, for example, in Nicolaou and co-workers' synthesis of rapamycin **12**⁵⁴ and Panek and Masse's synthesis of mycotrienin I and mycotrienol **92**⁵⁵ (Scheme 15), and in the previously discussed syntheses of myxalamide A **68**, viridenomycin **11**, erythrokyrine **63**, and the monomeric moiety of disorazole C₁ **76**. Duchene *et al.* have successfully applied Stille methodology to a variety of retinoids, including the trifluoromethyl retinoate **96**,⁵⁶ useful as a biological probe (Scheme 16). Their methodology uses enyne

Table 3

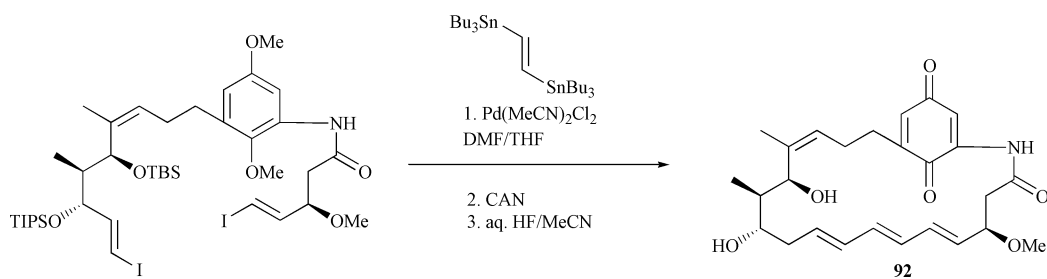
Core	Iodide	Retinoid ^a	Yield (%)
95a	(<i>E</i>)- 98	All- <i>trans</i> -9-Nor-retinoic acid	50
95a	(<i>Z</i>)- 98	(13 <i>Z</i>)-9-Nor-retinoic acid	45
95b	(<i>E</i>)- 98	All- <i>trans</i> retinoic acid	73
95b	(<i>Z</i>)- 98	(13 <i>Z</i>)-Retinoic acid	70

^a See Fig. 7 for retinoid numbering.

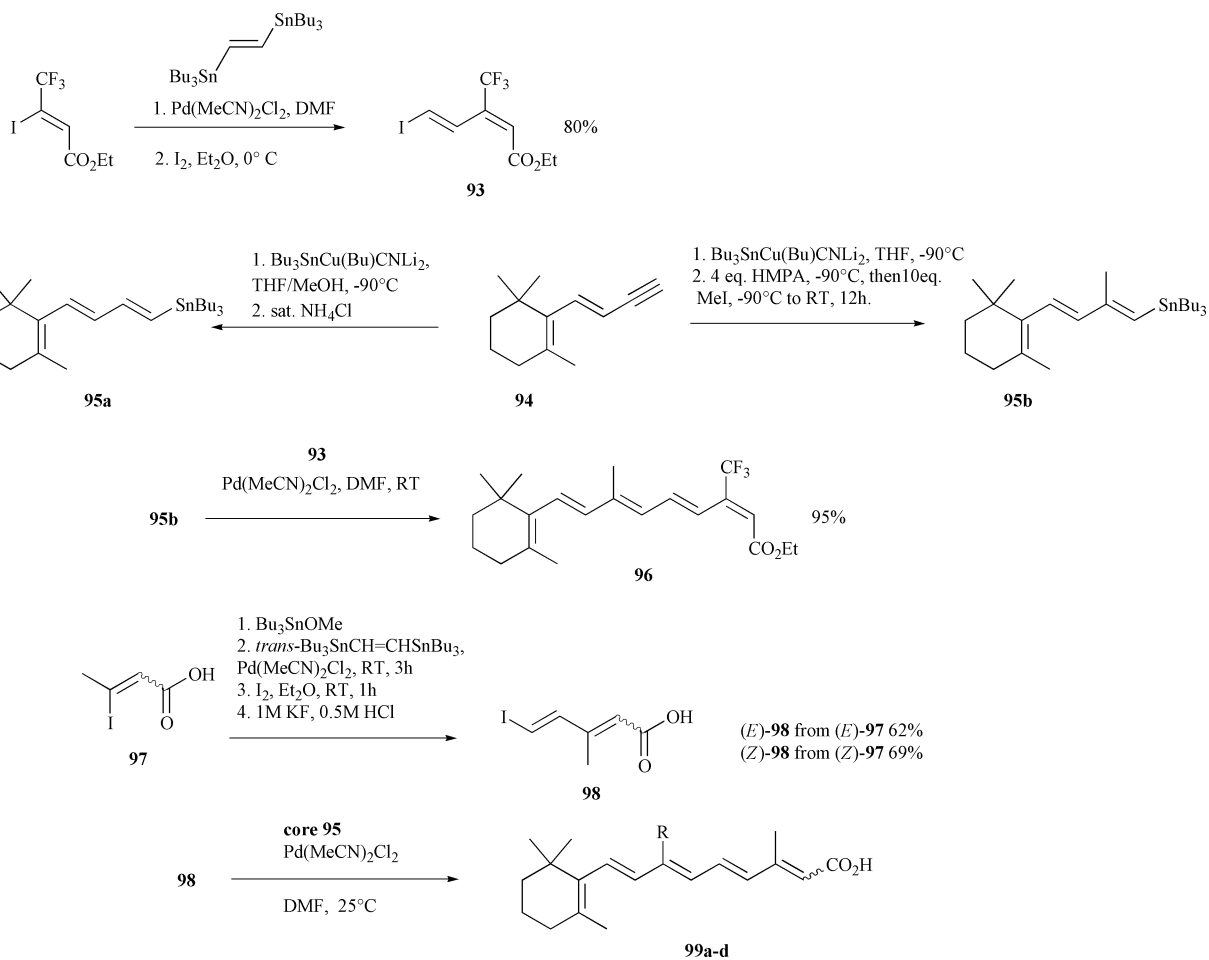


Scheme 14

precursor **94** derived from β-ionone,⁵⁷ which may either be converted into dienylstannane **95a** by treatment with the Lipshutz reagent⁵⁸ in the presence of methanol, or into the trisubstituted vinylstannane **95b** by trapping the intermediate vinylcuprate with methyl iodide. Stille couplings of β-iodovinyl acids **98** derived from tetrolic acid *via* iodides **97**, with stannanes **95** gives the retinoids stereoselectively (Table 3).⁵⁹ Fluorine substituted polyenes have traditionally been obtained *via* HWE or Julia reactions, but these methods are hampered by poor *E/Z* selectivity, particularly when forming the 9-*trans* trisubstituted double bond common in many retinoids. Yoshihara and co-workers have recently described a preparation of 9-*trans*-9-desmethyl-9-fluororetinal **101** and analogues based upon a four component coupling approach.⁶⁰ Due to the failure of the 4-hydroxybutynoate **100** to undergo fluorination under reported



Scheme 15



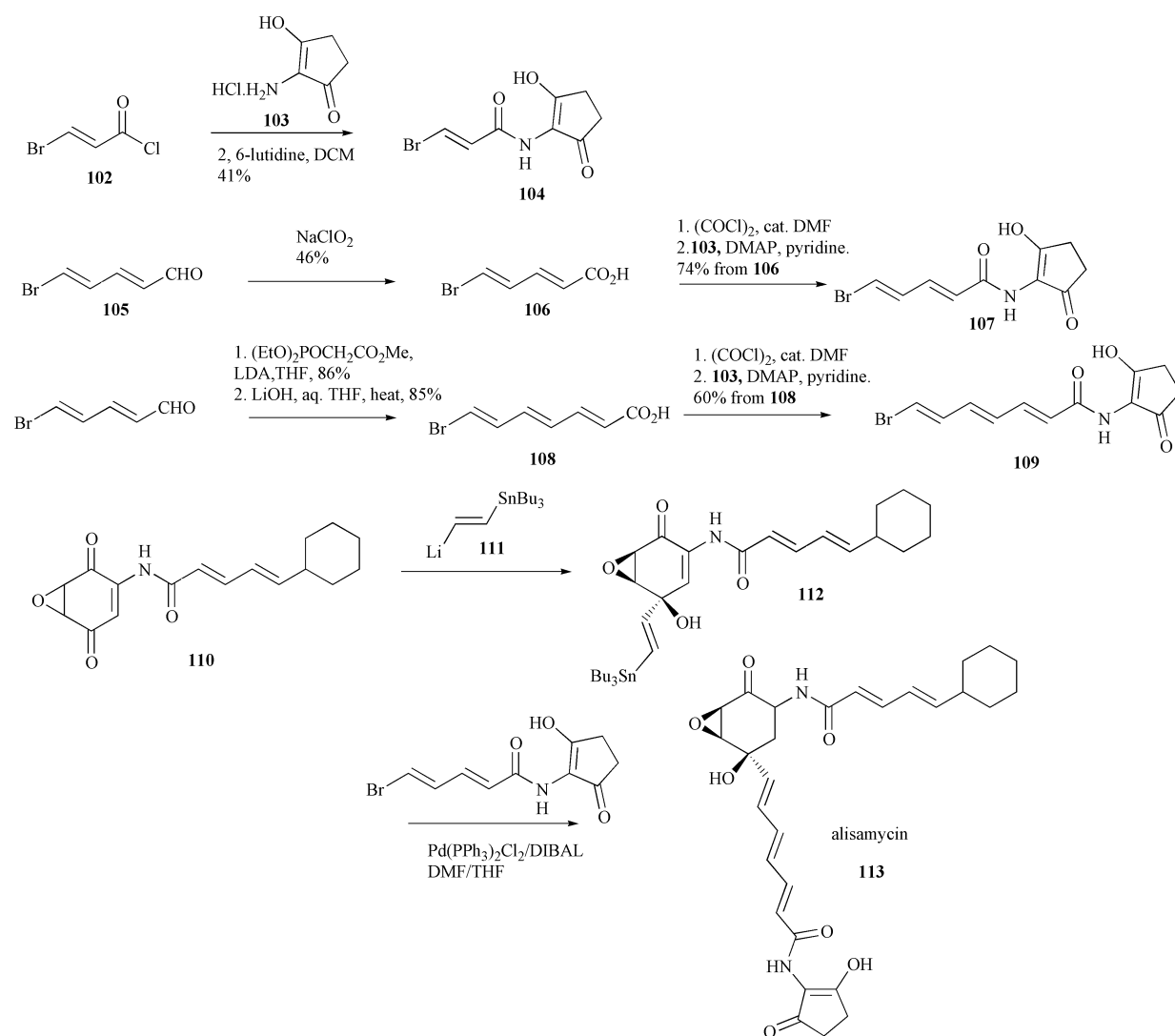
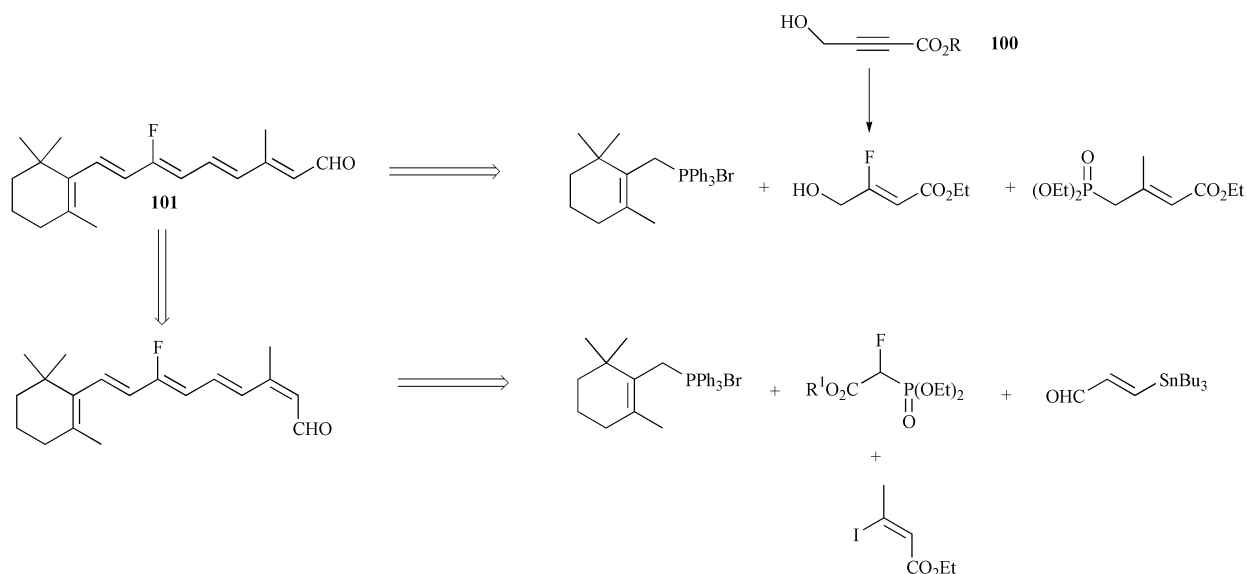
Scheme 16

conditions, the second approach utilizing Wittig, HWE and Stille chemistry was successfully adopted (Scheme 17). de Lera and co-workers have also exhaustively examined applications of the Stille reaction for the synthesis of retinoid skeletons. They studied various disconnections and then developed optimized approaches for each strategy, providing useful information regarding the influence of steric factors on the Stille reaction, as well as useful retinoid building blocks.⁶¹

The first complete synthesis of a manumycin antibiotic was achieved by Taylor and co-workers with alisamycin **113** (Scheme 18).⁶² Fundamental to this synthesis was the introduction of the unsaturated 2-amino-3-hydroxycyclopentenone forming the lower polyene chain. This moiety is present in many *Streptomyces* sp. metabolites, notably in many of the manumycin antibiotics. Taylor chose to insert this portion *via* the Stille reaction of 2-amino-3-hydroxycyclopentenone hydrohalides and vinylstannanes, thus treatment of known acyl chloride **102** with key amine hydrochloride **103** (prepared from cyclopentanedione) leads to vinyl bromide **104**. Higher vinylologues **107** and **109**, suitable for synthesizing the manumycins, can be prepared *via* HWE homologation of bromodienal **105**. Elaboration of the quinone **110** with the Corey-Wollenburg

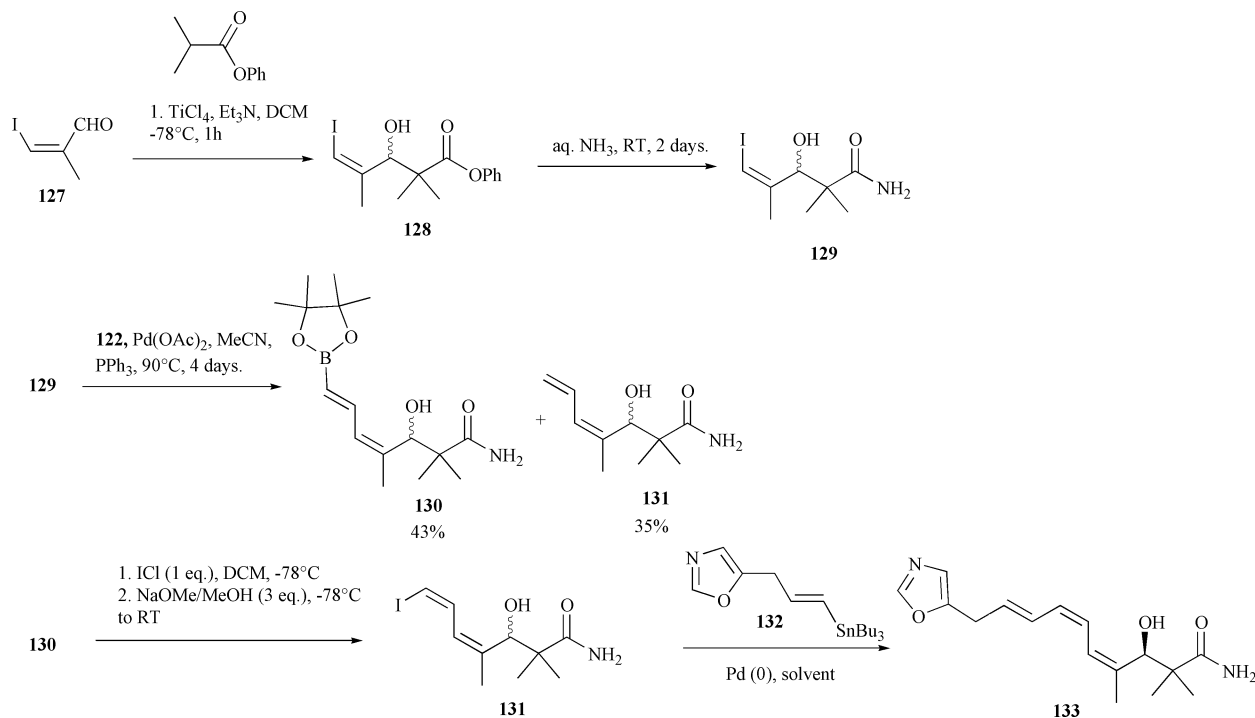
reagent⁶³ **111** gave a mixture of enantiomeric vinylstannanes; Stille coupling of these with bromodiene **107** led to alisamycin and *ent*-alisamycin. Taylor's group have successfully applied this strategy to a great number of the manumycin antibiotics and related polyene natural products.⁶⁴

Like the Stille coupling reaction, the closely related Suzuki reaction⁶⁵ has found utility in polyene synthesis. Earlier work towards polyene synthesis was carried out by Suzuki and co-workers, who successfully applied the reaction to the stereoselective synthesis of triene-containing natural products,⁶⁶ and later to the synthesis of trisporol B.⁶⁷ The Suzuki reaction can operate under extremely mild conditions making it an ideal way to synthesize potentially unstable polyenes. An example of this has already been seen in the total synthesis of myxalamide A, in which the synthesis was completed *via* Suzuki coupling of **69** and **70**.⁴⁶ de Lera's group have also presented a detailed study of highly stereoselective retinoid syntheses achieved *via* the thallium-accelerated Suzuki reaction carried out under ambient temperatures. These mild conditions were chosen to be compatible with the known instability of vitamin A and its derivatives. Their approach illustrates the great flexibility offered by the palladium coupling reactions, in that the choice of which



moiety to derive from the alkenyl iodide or alkenylboronic acid is dictated by relative ease of preparation. Preparation of retinol and its 9- and/or 11-desmethyl analogues (Scheme 19) was achieved through coupling of either boronic acids **116** with iodide **114** (also derived from enyne **94**), or with boronic acid **119** and iodides **117**, with total retention of the geometries

of the coupling partners. The group have also used Suzuki chemistry for the stereocontrolled preparation of 9-desmethyl-retinoids, useful for bio-organic studies into protein-chromophore steric interactions involved in bacteriorhodopsin photocycles,⁶⁸ and more recently have applied Suzuki chemistry to the synthesis of 7-*cis*-retinoids.⁶⁹



Scheme 22

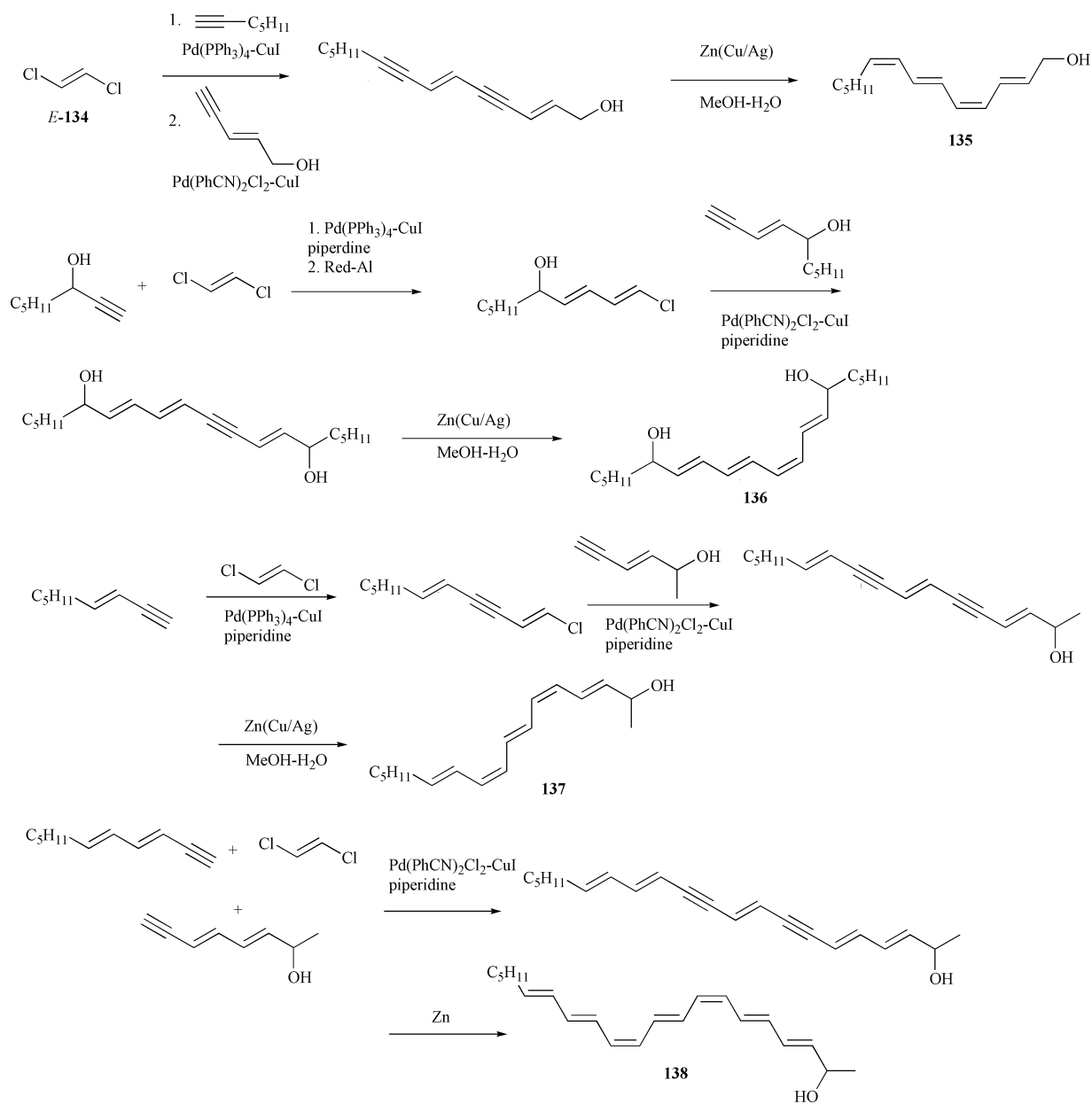
systems. Application of this methodology to the herbicidal agent phthoxazolin A is outlined in Scheme 22. Here, coupling of **122** and amide **129** (prepared from iodide **127** via an aldol reaction) demonstrates the functional group tolerance of these reactions, and gives the desired Heck product as the major product. Apart from this notable example, the Heck reaction, in its conventional guise, has found little utility in polyene synthesis. This is partially for the reason that, whilst the Stille reaction usually occurs with retention of stereochemical integrity of the coupling partners, the same cannot be said of the Heck reaction. Pattenden and co-workers recently compared the Heck and Stille reactions in his synthesis of pateamine A, a macrolide containing a (*Z,E*)-diene acrylate moiety in its macrocyclic core, and found the Stille reaction superior for this very reason.⁷⁷

In contrast, applications of the alkyne coupling variant of the Heck reaction towards polyene synthesis, the so-called Sonagashira coupling,⁷⁸ are numerous. Alami's group have devised techniques for the stereocontrolled synthesis of polyenes by sequential coupling of 1,2-dichloroethylenes **134** with a variety of acetylenes; selective reduction of the acetylene then leads to geometrically pure polyenes **135–138** (Scheme 23).⁷⁹ This methodology was applied to a short synthesis of lipoxin B **39** (Scheme 24). The same group have also developed a related protocol for accessing all-*E* polyenes. This protocol is based around α -chloro- ω -substituted hexatrienes **140**, obtained by two different approaches (Scheme 25). The first uses palladium catalyzed rearrangement of bis-allylic acetates, formed via bis-allylic alcohols **139**, whilst the second is based on the stereoselective reduction of homopropargylic alcohols **141** into (*E*)-homoallylic alcohols **142**, followed by elimination. Alcohols such as **142** may also be oxidized to give trienones and trienals, providing useful polyene building blocks having two reactive terminal functions, as the chlorine atom is able to participate in coupling reactions, with, for example, Grignard reagents, providing isomerically pure (*E,E,E*)-diarylhexatrienes **143**. The coupling partner may be acetylenic, as illustrated in a brief, high-yielding synthesis of navenone B **24**.⁸⁰

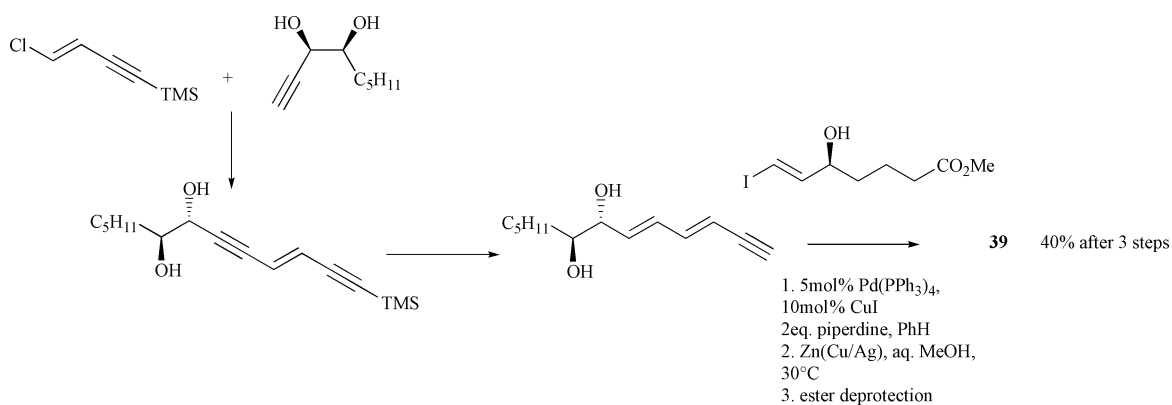
An obvious necessity for the palladium coupling reactions, when stereodefined polyenes are desired, is a stock of geometrically pure alkenyl halide or alkenylmetal. Many techniques giving access to these coupling partners have already been mentioned in this review.

Quintard and co-workers have done work on the regioselectivity of organotin cross-couplings, tuning the reaction conditions in order to obtain the requisite (*E,E*) dienylnit acetals **144** that may then be used to construct polyenic systems (Scheme 26).⁸¹ Uenishi's group have examined the stereoselective hydrogenolysis of 1,1-dibromoalkenes, providing access to (*Z*)-1-bromoalkenes **145** that are useful for (*E,Z*)-diene synthesis.⁸² Reduction of conjugated 1,1-dibromoalkenes affords 2-alkenyl or 2-alkynyl-substituted (*Z*)-1-bromo-1-alkenes **146**, useful synthons for polyene construction (Scheme 27). These may then undergo coupling with boronic acids, giving stereo-defined trienes such as **148**; coupling with alkynes is also possible. The validity of this approach to polyene synthesis was demonstrated by the synthesis of the unstable polyenes (11*Z*)-retinal and (2*Z,4E,6E*)-dehydrodendrolasin. The same group has also reported the nickel catalysed coupling of bromoalkenes **147** with Grignard reagents, giving potentially useful allylsilanes **150** when TMS-substituted Grignard reagents are employed, or alkylated 1,3-dienes e.g. **149** (Scheme 27).⁸³ Lipshutz and Lindsley have reported two highly specialized building blocks for the preparation of polyene systems. The first, stannylated dienyne **153**, is readily prepared through reaction of enal **151** with known Wittig salt **152** (Scheme 28). Dienenne **153** undergoes cross-coupling at the vinylstannane end with a variety of halides under different conditions, producing trienyne or tetraenyne products (Table 4).⁸⁴

Elaboration of the alkyne terminus of these products may be carried out either through initial hydrozirconation, transmetalation with aluminium, and trapping of the alane with a suitable electrophile, or via direct Negishi carboalumination followed by quenching. Both processes give rapid access to a variety of all-(*E*)-polyenes (Table 5). A synthesis of navenone B **24** is a noteworthy example. The method has further applications to natural products, for instance, preparation of the oxopentaene **156** common to the mycotoxins **154** and **155** (Scheme 29). Access to vinylogues of retinoic acid is also possible. Their redesigned trienyne (Scheme 30) **157** incorporates an inversion of polarity relative to **153** due to replacement of the stannyl diene moiety by vinylic bromide. **157** can thus undergo coupling with vinyl and dienylnit zinc reagents (Table 6). Elaboration of the alkyne terminus in the usual manner, either prior to, or after an initial coupling at the vinyl bromide



Scheme 23

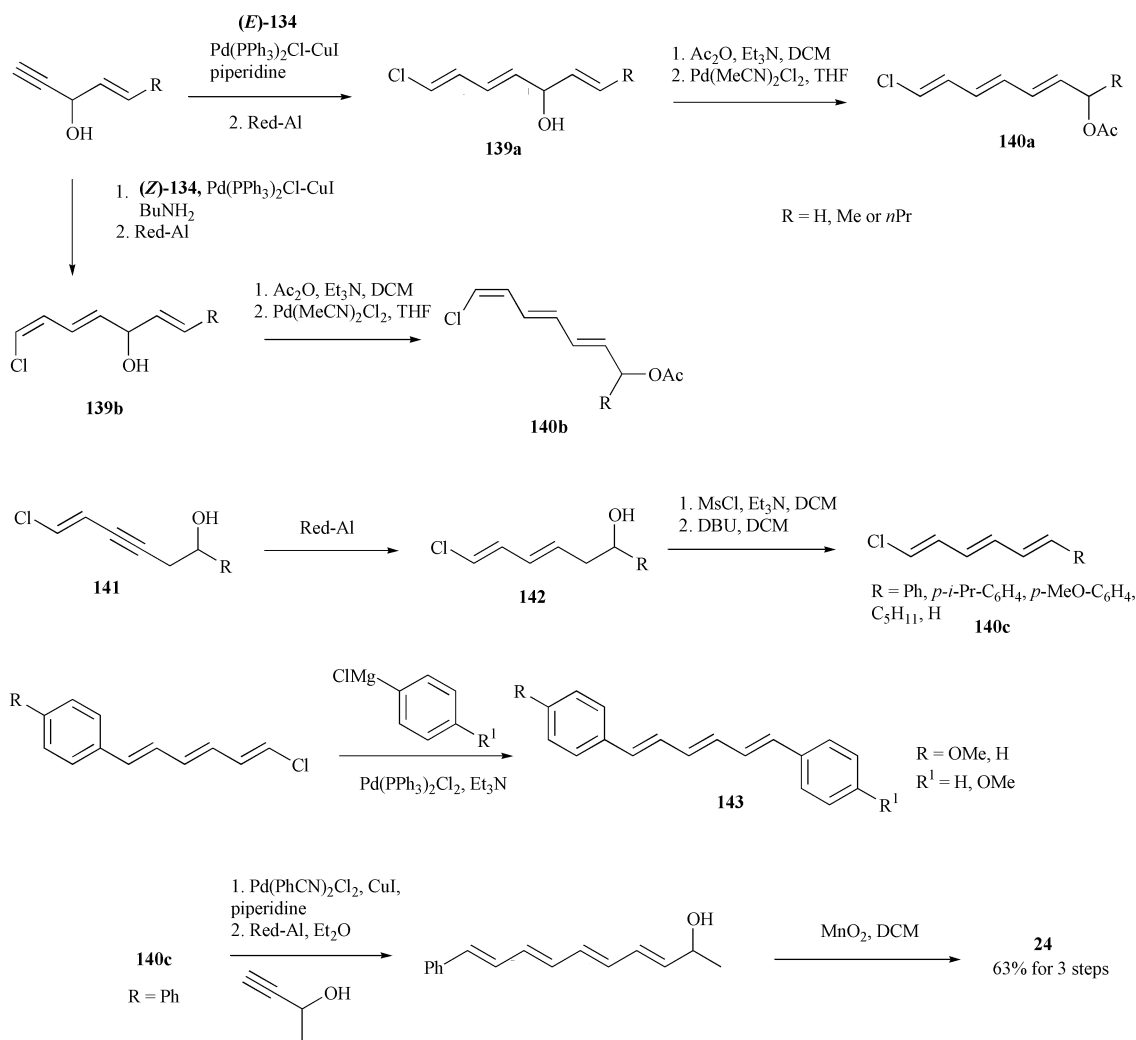


Scheme 24

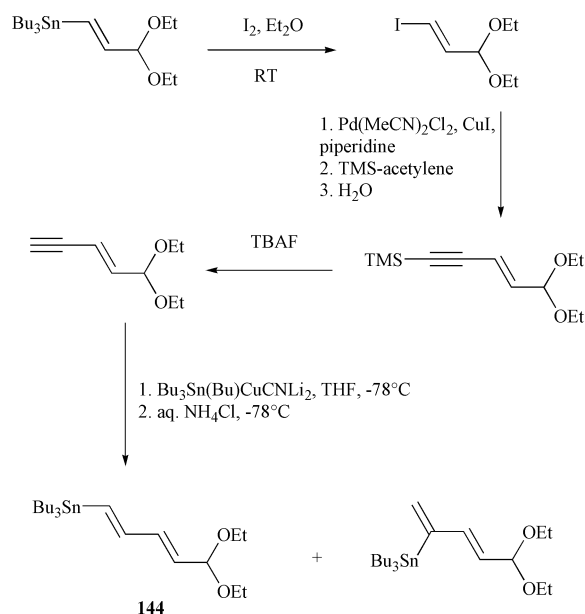
terminus, leads to oxopolyenes, and this approach was applied to a short synthesis of the oxohexaene **159**, common to the dermatostins, of which dermatostatin A **160** may be taken as a typical member (Scheme 31).⁸⁵

Zeng and Negishi have formulated a highly efficient and stereoselective procedure for the preparation of both sym-

metrical and unsymmetrical carotenoids. This procedure is based upon Zr-catalyzed carboalumination of conjugated oligoenynes followed by palladium and zinc catalyzed cross-coupling of the resultant alkenylalanes and allows ready access to members of the retinoid family. The procedure utilizes the two and four carbon synthons **162** and **163** respectively.



Scheme 25



Scheme 26

Accordingly, dihaloalkene **162** is readily converted to **163**; with these in hand the authors were able to prepare a variety of carotenoids *e.g.* β-carotene **165** was prepared in 41% overall yield in three linear steps from **94**. In a similar fashion γ-carotene **17** and vitamin A **33** were prepared in good yields (Scheme 32). It is noteworthy that in all cases, ≥99% stereo-

selectivity towards the drawn isomer was observed, leading to ≥99% isomerically pure polyenes.⁸⁶

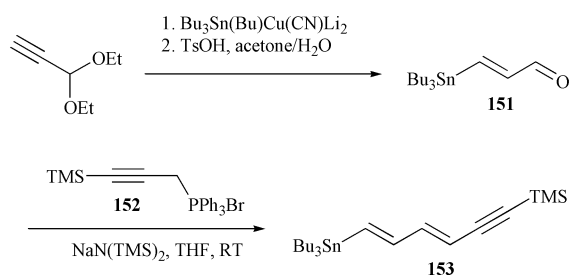
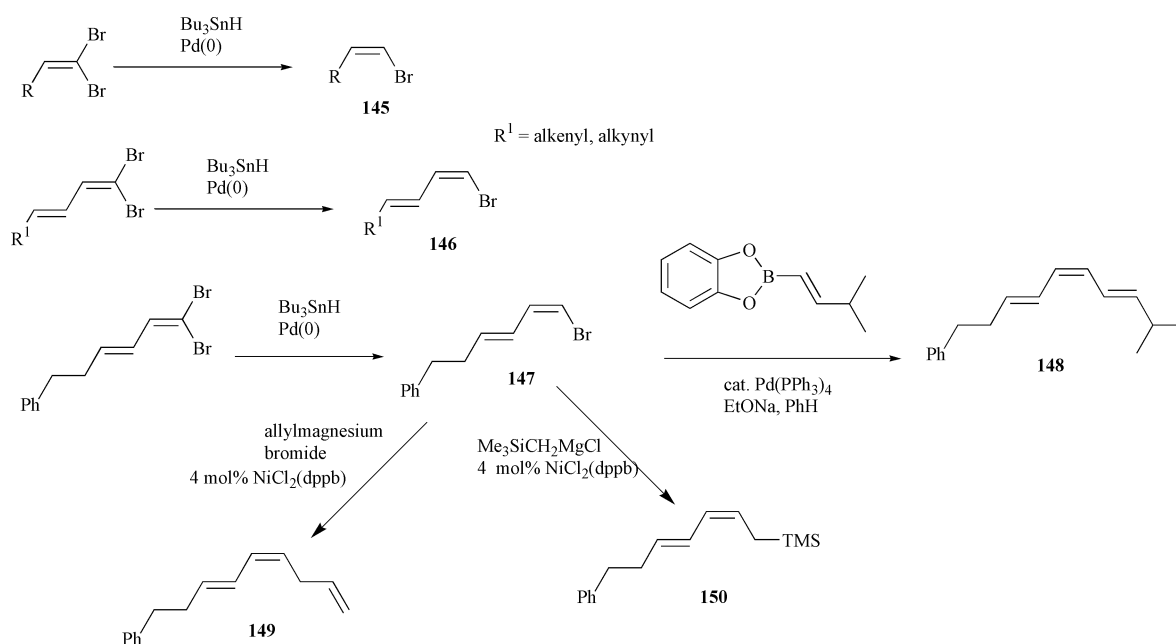
Wipf's group have prepared members of the manumycin family, using organozirconocene methodology to build the polyene sections.⁸⁷ This approach involves hydrozirconation of a functionalized alkyne *e.g.* **170**, followed by transmetalation with zinc, and *in situ* 1,2-addition of the organozinc reagent to α,β-unsaturated aldehydes. The approach is convergent and can be used to synthesize dienyl and trienyl side chains common to the manumycins, and facilitates analogue preparation. Scheme 33 exemplifies this protocol through the synthesis of the eastern side chain of asukamycin **171** and southern side-chain of nisamycin **172**.

Two recent applications of organoiron methodology towards polyene synthesis have been disclosed (Scheme 34). The first, aimed at stereoselective retinoid synthesis, uses a Peterson reaction between (β-ionylideneacetaldehyde)tricarbonyliron complex **173** and a range of trimethylsilyl derivatives to access retinoid precursors **174–177**. Iron complex **175** was subsequently transformed into 11*Z*-retinal **37** by conventional means.⁸⁸ The second application employs stoichiometric acyclic dienyl iron complexes. Accordingly, macrolactin A **178** was disconnected into fragments **179** and **181**, with a view to joining them together *via* nitrile oxide–olefin cycloaddition methodology. The (*E,Z*)-diene segment of **179** was prepared through addition of the strongly nucleophilic lithium anion of 2-(trimethylsilyl)ethylnitroacetate to the isolable *cisoid* cation **180**, giving an initial mixture of diastereomers that rearranges completely to the (*E,Z*) complex upon standing in chloroform.⁸⁹

Table 4

		method A, B or C	
		153 + halide	Product
Halide	Method	Product	Yield (%)
	A		83
	A		74
	A		80
	A		82
	B		72
	C		91

Method A: i). BuLi, THF, -78 °C; ii). ZnCl₂, 0 °C; iii). Pd(PPh₃)₄; iv). K₂CO₃, EtOH. Method B: i). 1.5 eq. CuCN, Pd₂(dba)₃, AsPh₃, NMP, 50 °C; ii). K₂CO₃, EtOH. Method C: i). 1.5 eq. CuCN, Pd₂(dba)₃, P(Fur)₃, NMP, 50 °C; ii). K₂CO₃, EtOH.



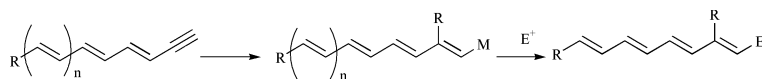
2.3 Miscellaneous methods

Several new methodologies have emerged recently for the preparation of polyenes. Naso and co-workers have successfully used bis(trimethylsilyl) derivatives to construct a variety of natural products, including the potent leukotriene B₃ methyl ester **186** (Scheme 35). Chemoselective acylation of bis(TMS) enyne **182** followed by enantioselective chemical reduction and catalytic hydrogenation of the triple bond leads to *E,Z*-diene **183** which is transformed into bromodiene **184**. A similar

sequence applied to bis(TMS) acetylene gives **185** which undergoes hydroboration with catecholborane to generate a vinylic boronic ester *in situ*; Suzuki–Miyaura coupling of this with **184** affords, after deprotection, **186** in 41% yield.⁹⁰

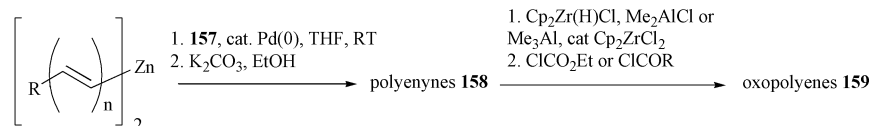
In a comparative study of several methods of polyene synthesis, including HWE strategies, an iterative strategy based upon the Wollenburg reagent **187**⁹¹ was found to be superior for the synthesis of the hexaene section of amphotericin B **4** (Scheme 36).⁹² This reagent is readily prepared and undergoes transmetalation to give either the lithiated species **188**, or Grignard-type reagents such as **191**, used recently by Rychnovsky's group to install four of the five double bonds of Filipin III **190** (Scheme 36).⁹³

Solladié's group have enjoyed much success using reductive elimination as a means of synthesizing polyenes in a stereoselective manner. Their strategy was recently exemplified by the first enantioselective synthesis of the alarm pheromone haminol-I **192** (Scheme 37). Diyne **194**, obtained through condensation of diacetylene with the appropriate aldehydes, yields the precursor dibenzoate diene **193** under standard conditions, with **192** attained as a single isomer following the reductive elimination step.⁹⁴

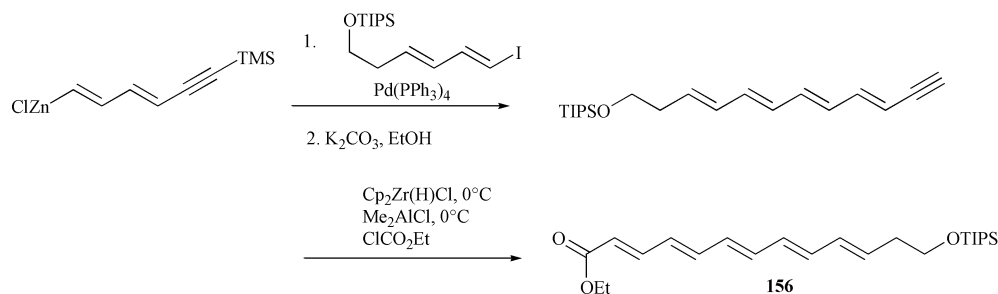
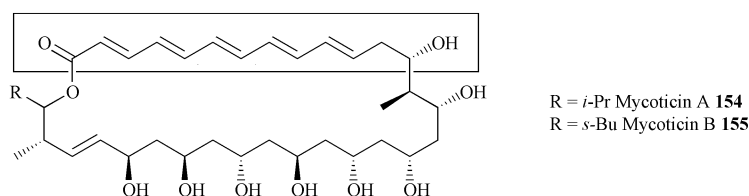
Table 5


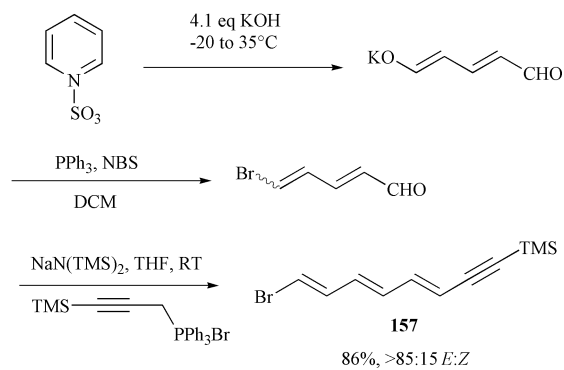
Substrate ^a	Yield (%)	Metallation/electrophile	Product	Yield (%)
	70-A	Cp ₂ Zr(H)Cl, DCM, Me ₂ AlCl, 0 °C, ClCO ₂ Et		71
	73-A	Cp ₂ Zr(H)Cl, DCM, Me ₂ AlCl, 0 °C, ClCOMe		82
	74-A	Cp ₂ Zr(H)Cl, DCM, Me ₂ AlCl, 0 °C, C ₃ H ₁₁ COCl		80
	75-A	Cp ₂ Zr(H)Cl, DCM, Me ₂ AlCl, 0 °C, ClCOMe		83
	91-B	Me ₃ Al, Cp ₂ Zr(H)Cl, ClCH ₂ CH ₂ Cl, ClCO ₂ CH ₂ CCl ₃		73

^a Prepared from **153** and the corresponding vinyl halide by methods A or B, cf. Table 4

Table 6


Polyenyne 158 (from 157)	Yield (%)	Metallation/electrophile	Oxopolyene 159 (from 158)	Yield (%)
	78	Cp ₂ Zr(H)Cl Me ₂ AlCl ClCO ₂ Et		68
	73	Cp ₂ Zr(H)Cl Me ₂ AlCl ClCO ₂ Et		73
	80	cat. Cp ₂ Zr ₂ Cl Me ₃ Al ClCO ₂ - <i>i</i> -Bu		75
	85	Cp ₂ Zr(H)Cl Me ₂ AlCl ClCOMe		70
	73	cat. Cp ₂ ZrCl ₂ Me ₃ Al ClCO ₂ Me		74

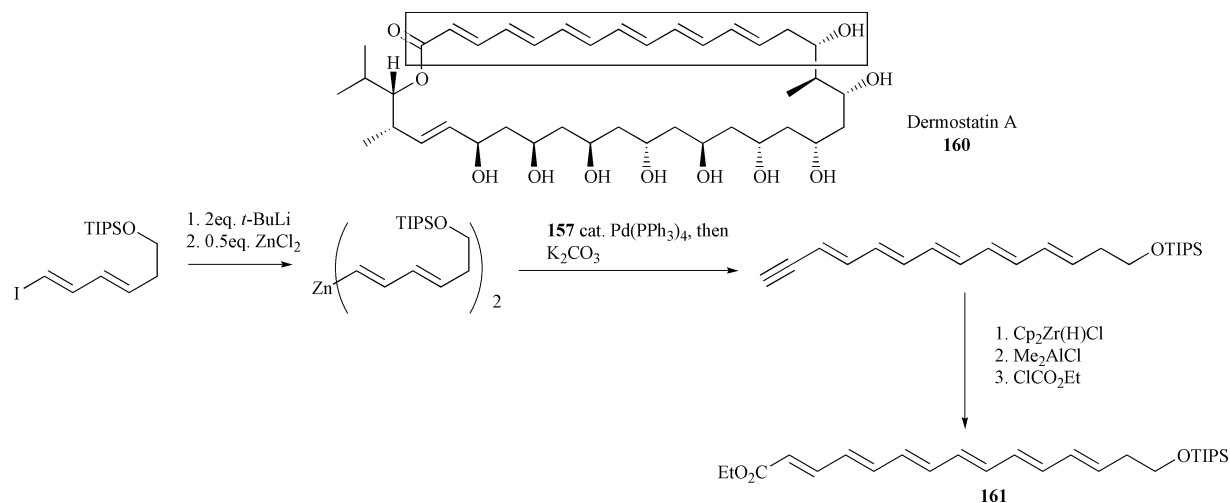

Scheme 29



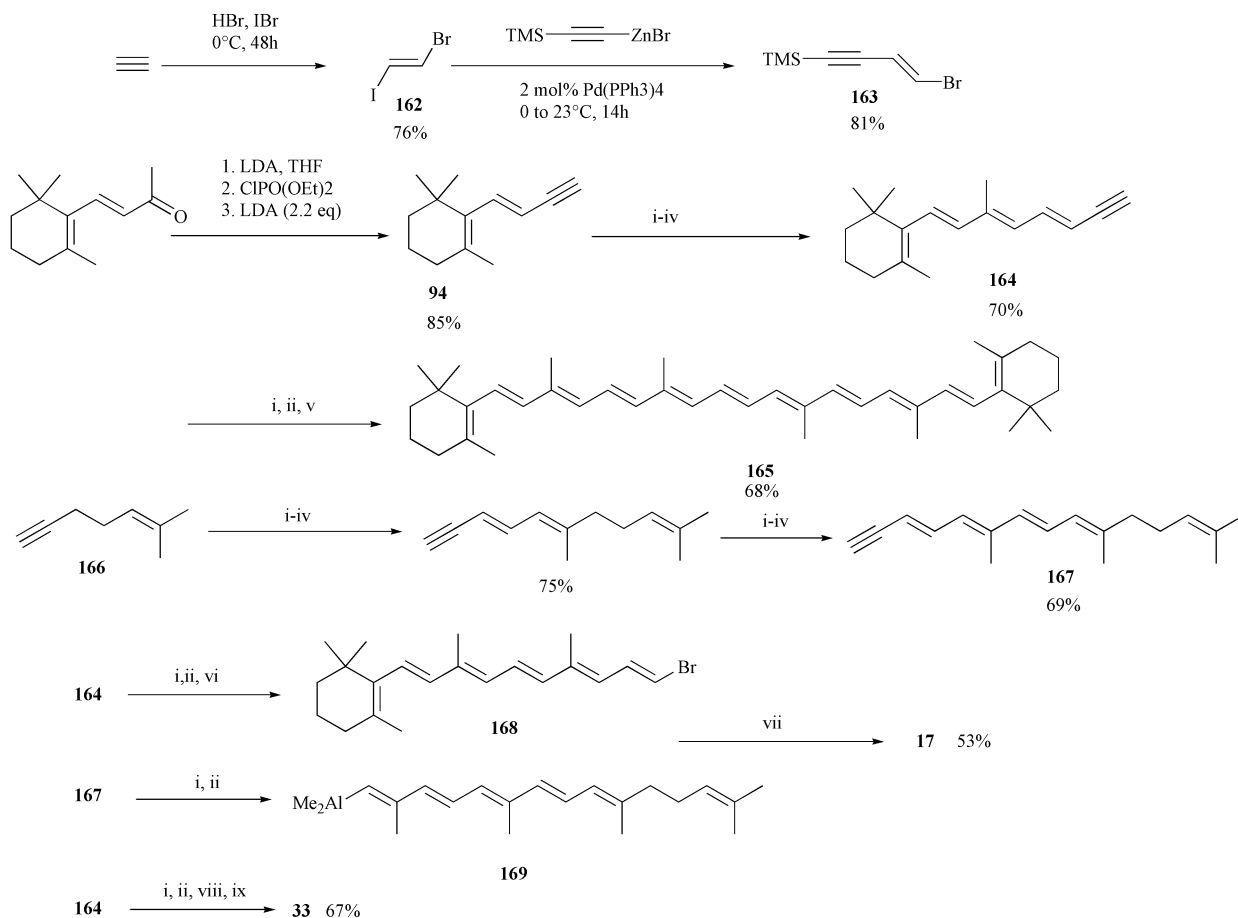
Scheme 30

A pericyclic approach to the 9-*cis*-retinoids was recently put forward by de Lera and co-workers (Scheme 38). A domino sequence, involving a reversible [2,3]-allyl sulfonate to allyl sulfoxide rearrangement (**196–197**), followed by propargylic sulfonate to vinylallenyl sulfoxide rearrangement (**199–200**), and finally, an irreversible, doubly stereoselective [1,5]-hydrogen shift to give **201**. Desulfuration and deprotection afford 9-*cis*-retinoids **36** and **203** (Scheme 38).⁹⁵

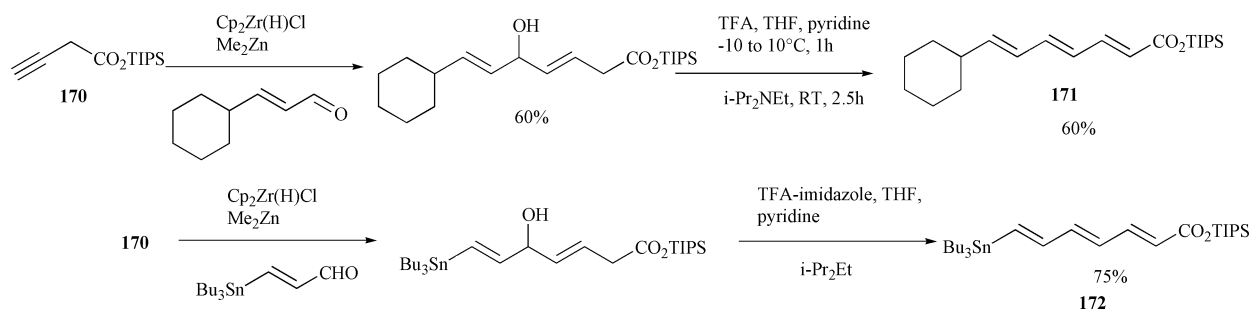
Taylor's group have used pyrylium salts to effect polyene synthesis; treatment of pyrylium tetrafluoroborate **204** with suitable organometallics allows access to a variety of retinoids, including the novel dehydro-demethyl-retinoid **206** (Scheme 39).⁹⁶



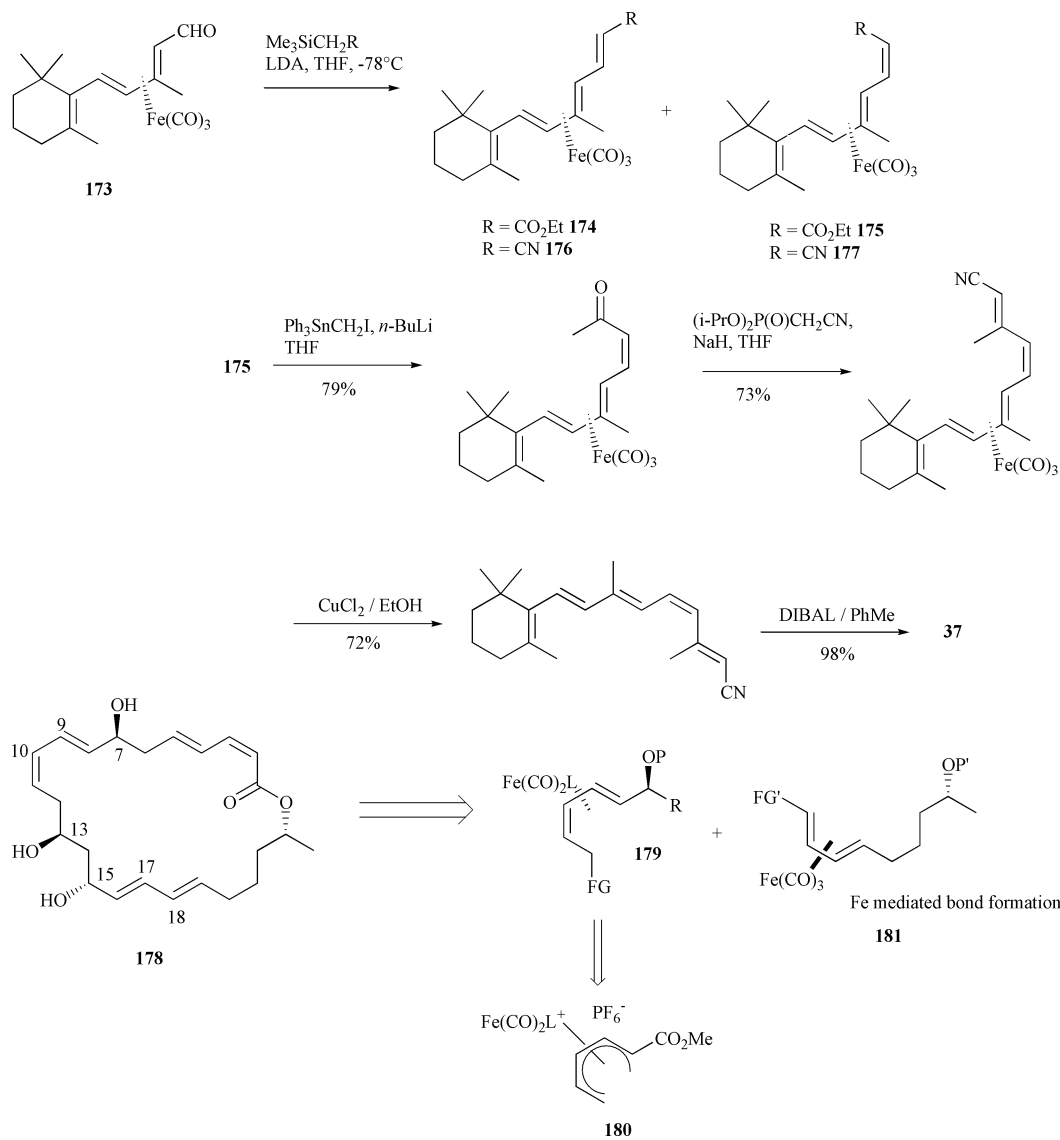
Scheme 31



Scheme 32 Conditions: i. Me_3Al (2 eq.), Cp_2ZrCl_2 (1 eq.) (CH_2Cl_2), 23°C , 4 h; ii. evaporation at 50°C and $<0.5 \text{ mmHg}$; iii. **163** (1.05 eq.), ZnCl_2 (1 eq.) in THF, 2.5 mol% $\text{Pd}_2(\text{dba})_3$, 10 mol% $\text{P}(\text{Fur})_3$, DMF, 23°C , 6 h; iv. K_2CO_3 , MeOH, 23°C , 3 h; v. **162** (0.5 eq.), ZnCl_2 (1 eq.) in THF, 2.5 mol% $\text{Pd}_2(\text{dba})_3$, 10 mol% $\text{P}(\text{Fur})_3$, DMF, 23°C , 8 h; vi. **162** (1.05 eq.), ZnBr_2 (1 eq.) in THF, 5 mol% $\text{Pd(PPh}_3)_4$, DMF, 23°C , 2 h; vii. ZnCl_2 (1 eq.) in THF, 2.5 mol% $\text{Pd}_2(\text{dba})_3$, 10 mol% $\text{P}(\text{Fur})_3$, DMF, 23°C , 6 h; viii. THF, *n*-BuLi (1 eq.), 23°C , 0.5 h; ix. $(\text{CH}_2\text{O})_n$, 23°C , 5 h. Fur = 2-furyl.



Scheme 33

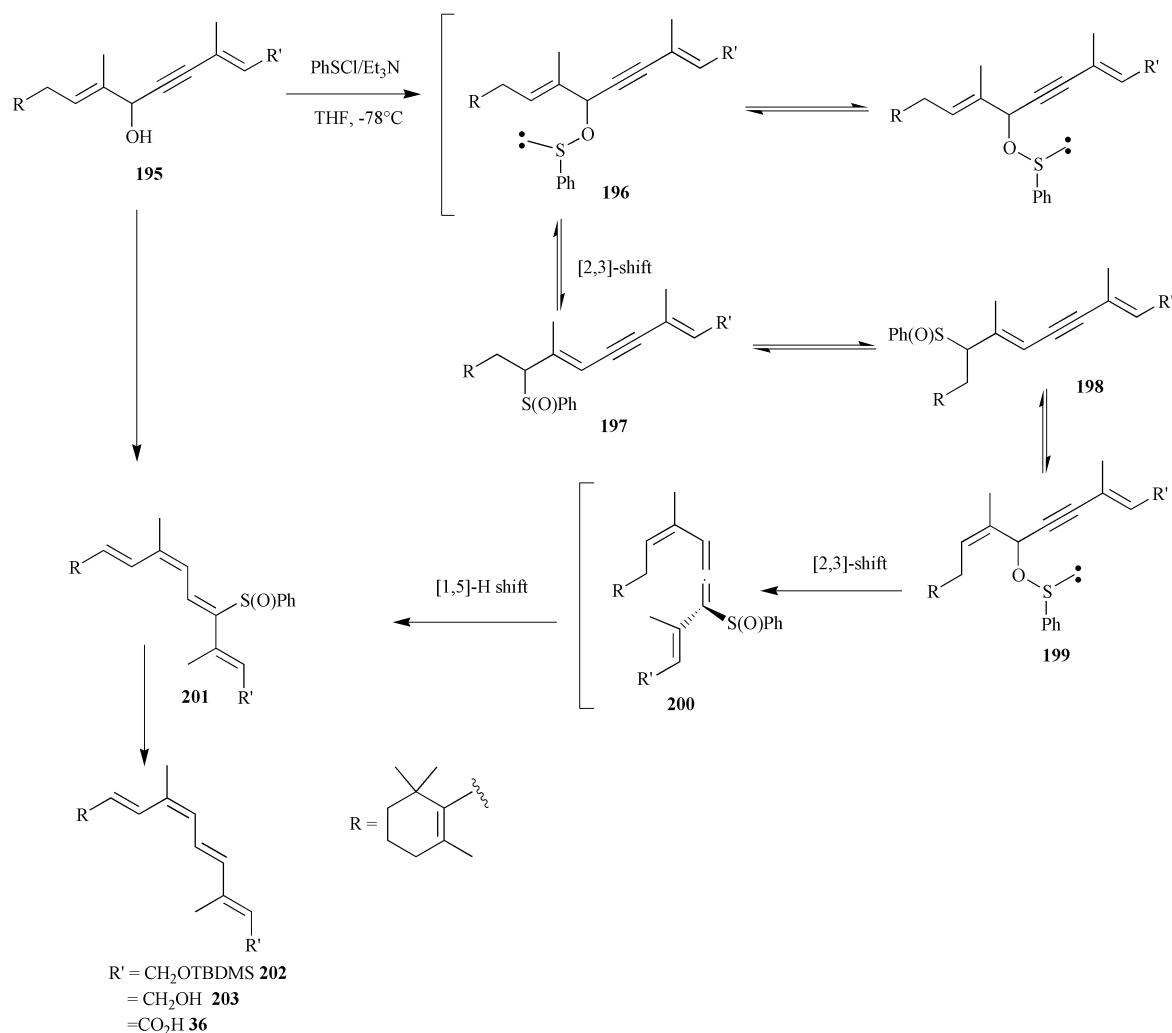


Scheme 34

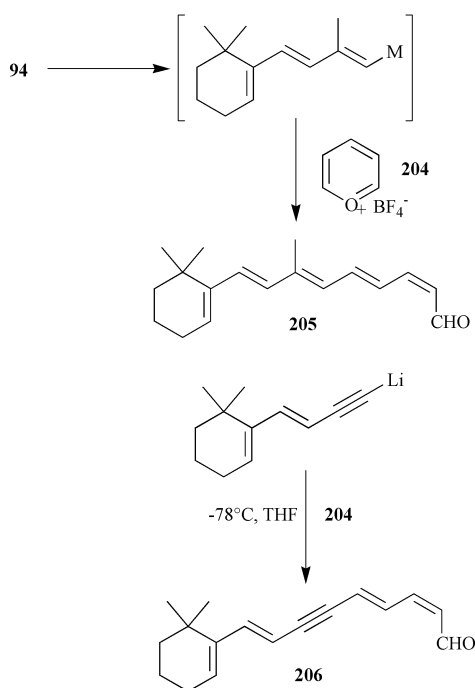
3 Conclusion

As can be seen from this review, much progress has been made in the area of polyene natural product synthesis over the last 30 or so years, and new methodology is constantly emerging. However, the ability to reliably construct polyenes with absolute control over alkene geometry, particu-

larly for those systems possessing *cis*-alkenyl units, remains an elusive goal. The increasing usage of polyene natural products in medicine, as drugs and as biological probes, as well as the general interest in polyenes for other applications, *e.g.* non-linear optics, should serve to fuel the continuing interest in this area, and help to achieve this goal.



Scheme 38



Scheme 39

4 References

- (a) *Leukotrienes and Lipoxygenase*, ed. J. Rockack, Elsevier, New York, 1989; (b) R. H. Green and P. F. Lambeth, *Tetrahedron*, 1983, **39**, 1687; (c) J. Evans, R. Zamboni, D. Nathaniel, C. Leveille and A. W. Ford-Hutchinson, *Prostaglandins*, 1985, **30**, 981; (d) R. J. Simmonds, *Chemistry of Biomolecules An Introduction*, Royal Society of Chemistry, Cambridge, 1992, ch. 7.
- (a) *The Retinoids*, ed. M. B. Born, A. B. Roberts and D. S. Goodman, Academic press, New York, 1984, vol. 1 and 2; (b) *The Retinoids: Biology, Chemistry and Medicine*, ed. M. B. Born, A. B. Roberts and D. S. Goodman, 2nd edn., Raven, New York, 1993; (c) *Chemistry and Biology of Synthetic Retinoids*, ed. M. L. Dawson and W. H. Okamura, CRC Press, Boca Raton, FL, 1990.
- (a) S. Omura and H. Tanaka, in *Macrolide Antibiotics: Chemistry, Biology and Practice*, ed. S. Omura, Academic Press, New York, 1984, pp. 351; (b) J. M. T. Hamilton-Miller, *Bacteriol. Rev.*, 1973, **37**, 166; (c) J. Kotler-Brajtburg, G. Medoff, G. S. Kobayashi, S. Boggs, D. Schlessinger, R. C. Pandey and K. L. Rinehart, *Antimicrob. Agents Chemother.*, 1979, **15**, 716; (d) J. A. Semlyen, *Large Ring Molecules*, Wiley, Chichester, 1996.
- S. D. Rychnovsky, *Chem. Rev.*, 1995, **95**, 2021.
- H. Maehr, R. Yang, L.-N. Hong, C.-M. Liu, M. H. Hatada and L. T. Todaro, *J. Org. Chem.*, 1989, **54**, 3816.
- J. M. Lancelin and J. M. Beau, *J. Am. Chem. Soc.*, 1990, **112**, 4060.
- G. J. McGarvey, J. A. Mathys and K. J. Wilson, *J. Org. Chem.*, 1996, **61**, 5704.
- R. W. Holz, in *Antibiotics*, ed. F. E. Hahn, Springer, New York, 1979, vol. 5, p. 313.
- (a) K. Furihata, Y. Natori, N. Otaki, T. Sasaki, H. Seto, A. Shimazu and M. Sugita, *J. Antibiot.*, 1982, **35**, 1460; (b) K. Furihata, Y. Natori, N. Otaki, T. Sasaki, H. Seto, A. Shimazu and M. Sugita, *J. Antibiot.*, 1982, **35**, 1467.
- K. Furihata, Y. Natori, N. Otaki, T. Sasaki, H. Seto, A. Shimazu and M. Sugita, *J. Antibiot.*, 1982, **35**, 1474.
- For a review, see (a) I. Sattler, R. Thierecke and A. Zeeck, *Nat. Prod. Rep.*, 1998, **15**, 221; (b) H. G. Floss, *Nat. Prod. Rep.*, 1997, **14**, 433.
- Isolation and properties: manumycin A: (a) F. Buzzetti, E. Gümman, R. Hütter, W. Keller-Schierlein, L. Neipp, V. Prelog and H. Zähner, *Pharm. Acta Helv.*, 1963, **38**, 871; (b) R. Thierecke,

- M. Stellwaag and A. Zeeck, *J. Antibiot.*, 1987, **40**, 1549; (c) A. Zeeck, K. Schröder, K. Frobel, R. Grote and R. Thiericke, *J. Antibiot.*, 1987, **40**, 1530; (d) A. Zeeck, K. Frobel, C. Heusel, K. Schröder and R. Thiericke, *J. Antibiot.*, 1987, **40**, 1541. Alisamycin; (e) C. M. M. Franco, R. Maurya, E. K. S. Vijayakumar, S. Chatterjee, J. Blumbach and B. N. Ganguli, *J. Antibiot.*, 1991, **44**, 1289; (f) S. Chatterjee, E. Vijayakumar, C. Franco, J. Blumbach and B. N. Ganguli, *J. Antibiot.*, 1993, **46**, 1027; (g) K. Hayashi, M. Nakagawa, T. Fujita and M. Nakayama, *Biosci. Biotech. Biochem.*, 1994, **58**, 1332. Asukamycin; (h) S. Omura, C. Kitao, H. Tanaka, R. Oiwa, Y. Takahashi, A. Nakagawa, M. Shimada and Y. Iwai, *J. Antibiot.*, 1976, **29**, 876; (i) K. Kakinuma, N. Ikekawa, A. Nakagawa and S. Omura, *J. Am. Chem. Soc.*, 1979, **101**, 3402; (j) H. G. Cho, I. Sattler, J. M. Beale, A. Zeeck and H. G. Floss, *J. Org. Chem.*, 1993, **58**, 7925. Nisamycin; (k) K. Hayashi, M. Nakagawa, T. Fujita, S. Tanimori and M. Nakayama, *J. Antibiot.*, 1993, **46**, 1904; (l) K. Hayashi, M. Nakagawa, T. Fujita, S. Tanimori and M. Nakayama, *J. Antibiot.*, 1994, **47**, 1110.
- 13 (a) R. Jansen, H. Irschik, H. Riechenbach, V. Wray and G. Hofle, *Liebigs Ann. Chem.*, 1994, 759; (b) R. Jansen, H. Irschik, H. Riechenbach, V. Wray and G. Hofle, *Tetrahedron Lett.*, 1985, **26**, 6031.
- 14 Isolation: (a) A. G. Andrews, S. Hertzberg, S. Liaaen-Jensen and M. P. Starr, *Acta Chem. Scand.*, 1973, **27**, 2383; A. G. Andrews, S. Hertzberg, S. Liaaen-Jensen and M. P. Starr, *Acta Chem. Scand.*, 1973, **27**, 2574; (b) A. G. Andrews, C. L. Jenkins, M. P. Starr, J. Shephard and H. Hope, *Tetrahedron Lett.*, 1976, **45**, 4023.
- 15 S. Sakuda, U. Gruce-Bigol, M. Itoh, T. Nishimura and Y. Yamada, *Tetrahedron Lett.*, 1995, **36**, 2777.
- 16 Isolation and characterization: (a) M. Nakagawa, K. Furihata, Y. Hayakawa and H. Seto, *Tetrahedron Lett.*, 1991, **32**, 659; (b) T. Hasegawa, T. Kamiya, T. Henmi, H. Iwasaki and S. Yamatodani, *J. Antibiot.*, 1975, **28**, 167.
- 17 Isolation and properties: (a) P. J. Belshaw, S. D. Meyer, D. D. Johnson, D. Romo, Y. Ikeda, M. Andrus, D. G. Alberg, L. W. Schultz, J. Clardy and S. L. Schreiber, *Synlett*, 1994, 381; (b) M. K. Rosen and S. L. Schreiber, *Angew. Chem., Int. Ed. Engl.*, 1992, **31**, 384; (c) S. L. Schreiber, *Science*, 1991, **251**, 283.
- 18 Isolation and properties: (a) M. C. McGowan, M. E. Callander and J. F. Lawlis, *Science*, 1951, **113**, 202; (b) J. H. Killough, G. B. Magill and R. C. Smith, *Science*, 1952, **115**, 71; (c) H. Katznelson and C. A. Jamieson, *Science*, 1952, **115**, 70.
- 19 For the first total synthesis, see E. J. Corey and B. B. Snider, *J. Am. Chem. Soc.*, 1972, **94**, 2549.
- 20 Isolation and properties: (a) S. Iwasaki, H. Kobayashi, J. Furukawa, M. Namikoshi and S. Okuda, *J. Antibiot.*, 1984, **32**, 354; (b) S. Iwasaki, M. Namikoshi, H. Kobayashi, J. Furukawa, S. Okuda, A. Itai, A. Kasuya, Y. Iitaka and Z. Sato, *J. Antibiot.*, 1986, **39**, 424; (c) C. Onozwa, M. Shimamura, S. Iwasaki and T. Oikawa, *J. Antibiot.*, 1984, **37**, 354; (d) H. L. McLeod, L. S. Murray, J. Wanders, A. Setanoians, M. A. Graham, N. Pavlidis, B. Heinrich, W. W. T. Huinink, D. J. T. Wagener, S. Aamdal and J. Verweij, *Br. J. Cancer*, 1996, **74**, 1944 and references cited therein.
- 21 (a) S. Iwasaki, M. Namikoshi, H. Kobayashi, J. Furukawa and S. Okuda, *Chem. Pharm. Bull.*, 1986, **34**, 1387; (b) S. Kiyoto, Y. Kawai, T. Kawakita, E. Kino, M. Okuhara, I. Uchida, H. Tanaka, M. Hashimoto, H. Terano, M. Kohsaka, H. Aoki and H. Imanaka, *J. Antibiot.*, 1986, **39**, 762.
- 22 (a) W. Steglich, *Pure Appl. Chem.*, 1989, **61**, 281; (b) I. Casser, B. Steffan and W. Steglich, *Angew. Chem., Int. Ed. Engl.*, 1987, **26**, 586.
- 23 B. Steffan, M. Praemassing and W. Steglich, *Tetrahedron Lett.*, 1987, **28**, 3667.
- 24 (a) *Carotenoids*, ed. O. Isler, Birkhauser Verlag, Basel, 1971, pp. 932; (b) *Carotenoid Chemistry and Biochemistry*, G. Britton and T. W. Goodwin, Pergamon Press, Oxford, 1982, pp. 224.
- 25 Isolation, characterization and properties: (a) G. Furstenberger and E. Hecker, *Experientia*, 1977, **33**, 986; (b) A. D. Kinghorn, *J. Nat. Prod.*, 1979, **42**, 112; (c) G. Furstenberger and E. Hecker, *Z. Naturforsch.*, 1985, **40**, 631; (d) T. Harayama, M. Kawanishi, S. Takabayashi and Y. Ito, *Cancer Lett.*, 1981, **12**, 175.
- 26 Compound **20**: M. Huang, *Phytochemistry*, 1990, **29**, 1317. Compound **21**: A. F. Barrero, A. Haidour, M. Muñoz-Dorado, M. Akssira, A. Sedqui and I. Mansour, *Phytochemistry*, 1998, **48**, 1237. Compound **22**: G. Solladié, C. Kalai, M. Adamy and F. Colobert, *Tetrahedron Lett.*, 1997, **38**, 6917 and references cited therein.
- 27 (a) H. L. Sleeper and W. Fenical, *J. Am. Chem. Soc.*, 1977, **99**, 2367; (b) W. Fenical, H. L. Sleeper, V. J. Paul, M. O. Stallard and H. H. Sun, *Pure Appl. Chem.*, 1979, **51**, 1865; (c) A. Spinella, L. A. Alvarez and G. Cimino, *Tetrahedron Lett.*, 1998, **39**, 2005.
- 28 (a) S. Matasunga, N. Fusetani and Y. Kato, *J. Am. Chem. Soc.*, 1991, **113**, 9690; (b) D. Wolf, F. J. Schmitz, F. Qiu and M. Kelly-Borges, *J. Nat. Prod.*, 1999, **62**, 170; (c) N. U. Sata, S. Matsunaga, N. Fusetani and R. W. M. Soest, *J. Nat. Prod.*, 1999, **62**, 969.
- 29 G. Hofle, S. Pohlan, G. Uhlig, K. Krabbe and D. Schumacher, *Angew. Chem., Int. Ed. Engl.*, 1994, **33**, 1495.
- 30 See, for example, (a) Y. Doi, M. Ishibashi, N. Yamaguchi and J. Kobayashi, *J. Nat. Prod.*, 1995, **58**, 1097 and references cited therein; (b) J. Kobayashi and M. Ishibashi, *Chem. Rev.*, 1993, **93**, 1753.
- 31 M. Murata, S. Matsuoaka, N. Matsumori, G. K. Paul and K. Tachibana, *J. Am. Chem. Soc.*, 1999, **121**, 870–871.
- 32 B. E. Maryanoff and A. B. Reitz, *Chem. Rev.*, 1989, **89**, 863.
- 33 (a) S. E. Kelly, in *Comprehensive Organic Synthesis*, ed. B. M. Trost and I. Fleming, Pergamon, Oxford, 1991, vol. 1, pp. 729–817; (b) B. J. Walker, in *Organophosphorus Reagents in Organic Synthesis*, ed. J. I. G. Cadogan, Academic Press, New York, 1979, p. 155.
- 34 C. Tode, Y. Yamano and M. Ito, *J. Chem. Soc., Perkin Trans. 1*, 1999, 1625.
- 35 S. Marumoto, H. Kogen and S. Naruto, *Tetrahedron*, 1999, **55**, 7145.
- 36 J. S. Yadav, D. K. Barma and D. Dutta, *Tetrahedron Lett.*, 1998, **39**, 143.
- 37 G. Pattenden and P. Patel, *J. Chem. Soc., Perkin Trans. 1*, 1991, **8**, 1941.
- 38 (a) K. C. Nicolaou and E. J. Sorensen, in *Classics in Total Synthesis: Targets, Strategies, Methods*, Wiley-VCH, 1996, ch. 24; (b) K. C. Nicolaou, T. K. Chakraborty, Y. Ogawa, R. A. Daines, N. S. Simpkins and G. T. Furst, *J. Am. Chem. Soc.*, 1988, **110**, 4660; (c) K. C. Nicolaou, R. A. Daines, J. Uenishi, W. S. Li, D. P. Papahatjis and T. K. Chakraborty, *J. Am. Chem. Soc.*, 1988, **110**, 4672; (d) K. C. Nicolaou, R. A. Daines, T. K. Chakraborty and Y. Ogawa, *J. Am. Chem. Soc.*, 1988, **110**, 4685; (e) K. C. Nicolaou, R. A. Daines, T. K. Chakraborty and Y. Ogawa, *J. Am. Chem. Soc.*, 1988, **110**, 4696.
- 39 Y. Mori, M. Asai, J. Kawade and H. Furukawa, *Tetrahedron*, 1995, **51**, 5315.
- 40 For synthetic efforts towards the rhizoxins, see (a) G. E. Keck, K. A. Savin, M. A. Weglarz and E. N. K. Cressman, *Tetrahedron Lett.*, 1996, **37**, 3291; (b) J. D. White, C. S. Nylund and N. J. Green, *Tetrahedron Lett.*, 1997, **38**, 7329; (c) J. D. White, M. A. Holoboski, C. S. Nylund and N. J. Green, *Tetrahedron Lett.*, 1997, **38**, 7333; (d) S. D. Burke, J. Hong and A. P. Mongin, *Tetrahedron Lett.*, 1998, **39**, 2239; (e) S. D. Burke, J. Hong, J. R. Lennox and A. P. Mongin, *J. Org. Chem.*, 1998, **63**, 6952; (f) A. S. Kende, B. E. Glass and J. R. Henry, *Tetrahedron Lett.*, 1995, **36**, 4741; (g) D. R. Williams, K. M. Werner and B. Feng, *Tetrahedron Lett.*, 1997, **38**, 6825.
- 41 J. A. Lafontaine, D. P. Provencal, C. Gardelli and J. W. Leahy, *Tetrahedron Lett.*, 1999, **40**, 4145.
- 42 Isolation and properties: (a) B. H. Howard and H. Raistrick, *Biochem. J.*, 1949, **44**, 227; (b) B. H. Howard and H. Raistrick, *Biochem. J.*, 1954, **57**, 212; (c) J. Shoji and S. Shibita, *Chem. Ind.*, 1964, 419; (d) J. Shoji, S. Shibata, U. Sanakawa, H. Taguchi and Y. Shibanuma, *Chem. Pharm. Bull.*, 1965, **13**, 1240.
- 43 D. J. Dixon, S. V. Ley, T. Gracza and P. Szolcsanyi, *J. Chem. Soc., Perkin Trans. 1*, 1999, **8**, 839.
- 44 M. Nazare and H. Waldmann, *Angew. Chem., Int. Ed.*, 2000, **39**, 1125.
- 45 Isolation and properties: (a) R. Jansen, G. Reifentahl, K. Gerth, H. Reichenbach and G. Höfle, *Liebigs Ann. Chem.*, 1983, 1081; (b) K. Gerth, R. Jansen, G. Reifentahl, G. Hofle, H. Irschik, B. Kunze, H. Reichenbach and G. Thierbach, *J. Antibiot.*, 1983, **36**, 1150; (c) R. Jansen, W. S. Sheldrick and G. Höfle, *Liebigs Ann. Chem.*, 1984, 78.
- 46 A. K. Mapp and C. H. Heathcock, *J. Org. Chem.*, 1999, **64**, 23.
- 47 G. Stork and K. Zhao, *Tetrahedron Lett.*, 1989, **30**, 2173.
- 48 M. C. Hillier, D. H. Park, A. T. Price, R. Ng and A. I. Meyers, *Tetrahedron Lett.*, 2000, **41**, 2821.
- 49 (a) A. W. Kruger and A. I. Meyers, *Tetrahedron Lett.*, 2001, **42**, 4301; (b) A. W. Kruger and A. I. Meyers, *Tetrahedron Lett.*, 2001, **42**, 4305.
- 50 R. Tamura, K. Saegusa, M. Kakihana and D. Oda, *J. Org. Chem.*, 1988, **53**, 2723.
- 51 W. C. Still and C. Gennari, *Tetrahedron Lett.*, 1983, **24**, 4405.
- 52 (a) X. Wei and R. J. K. Taylor, *Tetrahedron Lett.*, 1998, **39**, 3815; (b) L. Blackburn, X. Wei and R. J. K. Taylor, *Chem. Commun.*, 1999, 1337; (c) X. Wei and R. J. K. Taylor, *J. Org. Chem.*, 2000, **65**, 616.
- 53 (a) J. K. Stille, *Angew. Chem., Int. Ed. Engl.*, 1986, **25**, 508; (b) T. N. Mitchell, in *Metal-catalyzed cross coupling reactions*, ed. F. Diederich and P. J. Stang, Wiley-VCH, New York, 1998, pp. 167; (c) V. Farina, V. Krishnamurthy and W. J. Scott, *The Stille Reaction*, John Wiley & Sons, New York, 1998.
- 54 K. C. Nicolaou, T. K. Charkraborty, A. D. Piscopio, N. Minowa and P. Bertinato, *J. Am. Chem. Soc.*, 1993, **115**, 4419.
- 55 J. S. Panek and C. E. Masse, *J. Org. Chem.*, 1997, **62**, 8290.
- 56 J. Thibonnet, G. Prie, M. Abarbri, A. Duchene and J.-L. Parrain, *Tetrahedron Lett.*, 1999, **40**, 3151.

- 57 (a) E.-I. Negishi, A. O. King, W. L. Klima, W. Patterson and A. Silveira, Jr, *J. Org. Chem.*, 1980, **45**, 2526; (b) E.-I. Negishi, A. O. King and J. M. Tour, *Organic Synthesis*, 1990, **Coll. Vol. VII**, 63.
- 58 B. H. Lipshutz, E. L. Ellsworth, S. H. Dimock and D. C. Reuter, *Tetrahedron Lett.*, 1989, **30**, 2065.
- 59 J. Thibonnet, M. Abarbri, A. Duchêne and J.-L. Parrain, *Synlett.*, 1999, 141.
- 60 T. Shinada, N. Sekiya, N. Bojkova and K. Yoshida, *Tetrahedron*, 1999, **55**, 3675.
- 61 (a) B. Domínguez, B. Iglesias and A. R. de Lera, *Tetrahedron*, 1999, **55**, 15071; (b) B. Domínguez, B. Iglesias and A. R. de Lera, *J. Org. Chem.*, 1998, **63**, 4135.
- 62 (a) R. J. K. Taylor, L. Alcaez, I. Kapfer-Eyer, G. MacDonald, X. Wei and N. J. Lewis, *Synthesis*, 1998, 775; (b) G. MacDonald, L. Alcaez, N. Lewis and R. J. K. Taylor, *Tetrahedron Lett.*, 1998, **39**, 5433.
- 63 (a) E. J. Corey and R. H. Wollenburg, *J. Org. Chem.*, 1975, **40**, 3788; (b) E. J. Corey and R. H. Wollenburg, *J. Am. Chem. Soc.*, 1974, **96**, 5581.
- 64 (a) G. MacDonald, L. Alcaez, X. Wei, N. J. Lewis and R. J. K. Taylor, *Tetrahedron*, 1998, **54**, 9823; (b) J. J. Cronjé Grové, X. Wei and R. J. K. Taylor, *Chem. Commun.*, 1999, 421; (c) J. J. Cronjé Grové, X. Wei and R. J. K. Taylor, *J. Chem. Soc., Perkin Trans. 1*, 1999, 1143; (d) L. Alcaez, G. MacDonald, J. Ragot, N. J. Lewis and R. J. K. Taylor, *Tetrahedron*, 1999, **55**, 3707.
- 65 (a) A. Suzuki and N. Miyaura, *Chem. Rev.*, 1995, **95**, 2457; (b) A. Suzuki, in *Metal-catalyzed Cross-coupling reactions*, ed. F. Diederich and P. J. Stang, Wiley VCH, New York, 1998, p. 49.
- 66 N. Miyaura, H. Sugimoto and A. Suzuki, *Bull. Chem. Soc. Jpn.*, 1982, **55**, 2221.
- 67 N. Miyaura, Y. Satoh, S. Hara and A. Suzuki, *Bull. Chem. Soc. Jpn.*, 1986, **59**, 2029.
- 68 (a) R. Alvarez, B. Iglesias and A. R. de Lera, *Tetrahedron*, 1999, **55**, 13779; (b) R. Alvarez, B. Iglesias, S. López and A. R. de Lera, *Tetrahedron Lett.*, 1998, **39**, 5659.
- 69 R. Alvarez, B. Domínguez and A. R. de Lera, *Synth. Commun.*, 2001, **31**, 2083.
- 70 (a) Y. Kobayashi, T. Shimazaki and F. Sato, *Tetrahedron Lett.*, 1987, **28**, 5849; (b) Y. Kobayashi, T. Shimazaki, H. Taguchi and F. Sato, *J. Org. Chem.*, 1990, **55**, 5324.
- 71 (a) K. C. Nicolaou, J. Y. Ramphal, J. M. Palazon and R. A. Spanevello, *Angew. Chem., Int. Ed. Engl.*, 1989, **28**, 587; (b) K. C. Nicolaou, J. Y. Ramphal, N. A. Petasis and C. N. Serhan, *Angew. Chem. Int. Ed. Engl.*, 1991, **30**, 1100.
- 72 M. R. Reeder and A. I. Meyers, *Tetrahedron Lett.*, 1999, **40**, 3115.
- 73 (a) N. Hénaff and A. Whiting, *Org. Lett.*, 1999, **1**, 1137; (b) N. Hénaff and A. Whiting, *Tetrahedron*, 2000, **56**, 5193.
- 74 (a) A. R. Hunt, S. K. Stewart and A. Whiting, *Tetrahedron Lett.*, 1993, **34**, 3599; (b) S. K. Stewart and A. Whiting, *J. Organomet. Chem.*, 1994, **482**, 293; (c) S. K. Stewart and A. Whiting, *Tetrahedron Lett.*, 1995, **36**, 3925.
- 75 (a) H. C. Brown, T. Hamaoka and N. Ravindran, *J. Am. Chem. Soc.*, 1973, **95**, 5786; (b) H. C. Brown and J. B. Campbell, *J. Org. Chem.*, 1980, **45**, 389.
- 76 S. K. Stewart and A. Whiting, *Tetrahedron Lett.*, 1995, **36**, 3929.
- 77 (a) M. J. Remuñán and G. Pattenden, *Tetrahedron Lett.*, 2000, **41**, 7367; (b) C. Cálina and G. Pattenden, *Synlett.*, 2000, **11**, 1661.
- 78 (a) K. Sonogashira, in *Comprehensive Organic Synthesis*, ed. B. M. Trost and I. Fleming, Pergamon Press, New York, 1991, vol. 3, p. 481; (b) K. Sonogashira, Y. Tohda and N. Hagihara, *Tetrahedron Lett.*, 1975, **16**, 4467; (c) R. Rossi, A. Carpita and F. Bellina, *Org. Prep. Proced. Int.*, 1995, **27**, 129.
- 79 For a review see: (a) G. Linstrumelle and M. Alami, (*E*) and (*Z*)-Dichloroethylene, in *Encyclopedia of Reagents for Organic Synthesis*, ed. L. Paquette, Wiley, Chichester, 1995, vol. 3, 1710; (b) B. Crousse, M. Alami and G. Linstrumelle, *Tetrahedron Lett.*, 1995, **36**, 4245.
- 80 (a) M. Mladenova, M. Alami and G. Linstrumelle, *Tetrahedron Lett.*, 1996, **37**, 6547; (b) B. Crousse, M. Alami and G. Linstrumelle, *Tetrahedron Lett.*, 1997, **38**, 5297; (c) B. Crousse, M. Mladenova, P. Ducept, M. Alami and G. Linstrumelle, *Tetrahedron*, 1999, **55**, 4353.
- 81 V. Launay, I. Beaudet and J.-P. Quintard, *Bull. Soc. Chim. Fr.*, 1997, **134**, 937.
- 82 J. Uenishi, R. Kawahama and O. Yonemitsu, *J. Org. Chem.*, 1998, **63**, 8965.
- 83 J. Uenishi, R. Kawahama, Y. Izaki and O. Yonemitsu, *Tetrahedron*, 2000, **56**, 3493.
- 84 B. H. Lipshutz and C. Lindsley, *J. Am. Chem. Soc.*, 1997, **119**, 4555.
- 85 B. H. Lipshutz, B. Ullman, C. Lindsley, S. Pecchi, D. J. Buzard and D. Dickson, *J. Org. Chem.*, 1998, **63**, 6092.
- 86 F. Zeng and E.-I. Negishi, *Org. Lett.*, 2001, **3**, 719.
- 87 (a) P. Wipf and Y. Kim, *J. Org. Chem.*, 1994, **59**, 3518; (b) P. Wipf, Y. Kim and H. Jahn, *Synthesis*, 1995, 1549; (c) P. Wipf, W. Xu, H. Takahashi, H. Jahn and P. D. G. Coish, *Pure Appl. Chem.*, 1997, **69**, 639; (d) P. Wipf and P. D. G. Coish, *Tetrahedron Lett.*, 1997, **38**, 5073; (e) P. Wipf and P. D. G. Coish, *J. Org. Chem.*, 1999, **64**, 5053.
- 88 A. Wada, N. Fujioka, Y. Tanaka and M. Ito, *J. Org. Chem.*, 2000, **65**, 2438.
- 89 H. Bärmann, V. Prahlad, C. Tao, Y. K. Yun, Z. Wang and W. A. Donaldson, *Tetrahedron*, 2000, **56**, 2283.
- 90 F. Barbudri, V. Fiandanese, O. Hassan, A. Punzi and F. Naso, *Tetrahedron*, 1998, **54**, 4327 and references cited therein.
- 91 R. H. Wollenburg, K. F. Alibizati and R. J. Peries, *J. Am. Chem. Soc.*, 1977, **99**, 7365.
- 92 J. M. Williams and G. J. McGarvey, *Tetrahedron Lett.*, 1985, **26**, 4891.
- 93 T. I. Richardson and S. D. Rychnovsky, *Tetrahedron*, 1999, **55**, 8977.
- 94 G. Solladié, F. Somny and F. Colobert, *Tetrahedron: Asymmetry*, 1997, **8**, 801 and references cited therein.
- 95 B. Iglesias, A. Torrado and A. R. de Lera, *J. Org. Chem.*, 2000, **65**, 2696.
- 96 R. J. K. Taylor, K. Hemming and E. F. De Medeiros, *J. Chem. Soc., Perkin Trans. 1*, 1995, 2385.